

LABORATORY INVESTIGATION



journal homepage: https://laboratoryinvestigation.org/

# **Review Article**

# Recent Advances in Enterovirus A71 Infection and Antiviral Agents

Yanhong Wei<sup>a</sup>, Huihui Liu<sup>a</sup>, Da Hu<sup>a</sup>, Qun He<sup>a</sup>, Chenguang Yao<sup>a</sup>, Hanluo Li<sup>a</sup>, Kanghong Hu<sup>a,\*</sup>, Jun Wang<sup>b,\*</sup>

<sup>a</sup> National "111" Center for Cellular Regulation and Molecular Pharmaceutics, Key Laboratory of Fermentation Engineering (Ministry of Education), Hubei Provincial Cooperative Innovation Center of Industrial Fermentation, Hubei Key Laboratory of Industrial Microbiology, Sino-German Biomedical Center, Hubei University of Technology, Wuhan, China; <sup>b</sup> Department of Pathology, Tongji Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan, China

# A R T I C L E I N F O

Article history: Received 17 May 2023 Revised 29 October 2023 Accepted 20 November 2023

*Keywords:* enterovirus A71 antiviral agents receptor viral replication signal pathway

# ABSTRACT

Enterovirus A71 (EV-A71) is one of the major causative agents of hand, foot, and mouth disease (HFMD) that majorly affects children. Most of the time, HFMD is a mild disease but can progress to severe complications, such as meningitis, brain stem encephalitis, acute flaccid paralysis, and even death. HFMD caused by EV-A71 has emerged as an acutely infectious disease of highly pathogenic potential in the Asia-Pacific region. In this review, we introduced the properties and life cycle of EV-A71, and the pathogenesis and the pathophysiology of EV-A71 infection, including tissue tropism and host range of virus infection, the diseases caused by the virus, as well as the genes and host cell immune mechanisms of major diseases caused by enterovirus 71 (EV-A71) infection, such as encephalitis and neurologic pulmonary edema. At the same time, clinicopathologic characteristics of EV-A71 infection were introduced. There is currently no specific medication for EV-A71 infection, highlighting the urgency and significance of developing suitable anti-EV-A71 agents. This overview also summarizes the targets of existing anti-EV-A71 agents, including virus entry, translation, polyprotein processing, replication, assembly and release; interferons; interleukins; the mitogen-activated protein kinase, phosphatidylinositol 3-kinase, and protein kinase B signaling pathways; the oxidative stress pathway; the ubiquitin-proteasome system; and so on. Furthermore, it overviews the effects of natural products, monoclonal antibodies, and RNA interference against EV-A71. It also discusses issues limiting the research of antiviral drugs. This review is a systematic and comprehensive summary of the mechanism and pathological characteristics of EV-A71 infection, the latest progress of existing anti-EV-A71 agents. It would provide better understanding and guidance for the research and application of EV-A71 infection and antiviral inhibitors.

© 2023 United States & Canadian Academy of Pathology. Published by Elsevier Inc. All rights reserved.

# Introduction

Human enteroviruses are members of the genus *Enterovirus* in the *Picornaviridae* family, which comprises a group of small,

\* Corresponding authors.

nonenveloped, positive-sense single-stranded RNA viruses. Enterovirus A71 (EV-A71) belongs to the human enterovirus species. EV-A71 can cause hand, foot, and mouth disease (HFMD) among children aged <5 years owing to their underdeveloped immune systems, according to the Chinese guidelines for diagnosing and treating HFMD.<sup>1</sup> Its clinical manifestation is milder, usually with fever, oral ulcers, and skin rashes on the hands and feet. However, HFMD caused by EV-A71 infection sometimes accompanies severe neurologic complications (Fig. 1), such as



0023-6837/\$ - see front matter © 2023 United States & Canadian Academy of Pathology. Published by Elsevier Inc. All rights reserved. https://doi.org/10.1016/j.labinv.2023.100298

E-mail addresses: hukh@hbut.edu.cn (K. Hu), wangjun2028@whu.edu.cn (J. Wang).

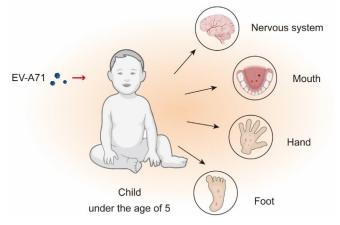


Figure 1.

Clinical manifestations of EV-A71 infection. The clinical features of EV-A71 are oral ulcers and skin rashes on the hands and feet. EV-A71 infection sometimes causes severe neurologic disorders, such as aseptic meningitis and encephalitis. EV-A71, enterovirus A71.

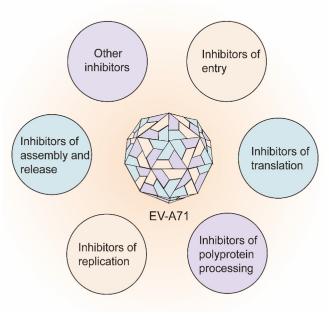
aseptic meningitis, encephalitis, poliomyelitis-like paralysis, and even death.

In 1969, enterovirus 71 (EV-A71) was detected in the United States in a 9-month baby with encephalitis. The virus was also identified in California,<sup>2</sup> and after that, it spread across the world, especially in the Asia-Pacific region,<sup>3-7</sup> including China,<sup>6,7</sup> Singapore,<sup>8</sup> Vietnam,<sup>9</sup> Japan,<sup>10</sup> and so on. EV-A71 caused 126 deaths in 2008 in China,<sup>11</sup> and the largest outbreak in China occurred in 2010, with an estimated 1.7 million infections, of which 27,000 caused severe neurologic complications, resulting in 905 fatalities.<sup>12</sup> Fatal cases were also reported in Asia as recently as 2012.<sup>13</sup>

EV-A71 produces a broad spectrum of clinical manifestations. The majority of infected individuals have asymptomatic infection. Mild cases are characterized as cutaneous diseases, such as HFMD and herpangina. Occasionally, EV-A71 infection can cause serious neurologic diseases, even life-threatening. Currently, EV-A71 vaccines are available on the Chinese market. However, owing to the mutating nature of RNA viruses, these vaccines may not provide long-term protection. Moreover, the vaccine mainly plays a preventive role, and the therapeutic drugs specific to EV-A71 in clinical use are still blank.<sup>14</sup> Therefore, there is an urgent need for anti-EV-A71 drugs to combat HFMD. This review summarized the pathological mechanisms caused by EV71 infection and focuses on the whole process of EV-A71 infection and related host factors, and classifies antiviral agents targeting EV-A71 into 6 major categories, namely inhibitors of entry, translation, polyprotein processing, replication, assembly, and release, and other inhibitors (Fig. 2).

#### The Properties and Life Cycle of the Virus

The EV-A71 virus is nonenveloped and icosahedral with a diameter of approximately 20 to 30 nm and a single-stranded positive polarity (+) RNA genome. The life cycle of EV-A71 (Fig. 3) starts with binding to one or more receptors on the cell surface. When EV-A71 binds to receptors, structural changes occur in the viral capsid and cell membrane. Then, partial pores are formed in the cell membrane to facilitate the penetration of the virion's positive-strand (+) RNA genome into the cytoplasm. The viral protein VPg (3B) acts as a primer for viral replication and



#### Figure 2.

Classification of antiviral agents for enterovirus A71 (EV-A71) infection. There are 6 major categories of anti-EV-A71 agents, namely inhibitors of entry, translation, polyprotein processing, replication, assembly and release, and other EV-A71 inhibitors.

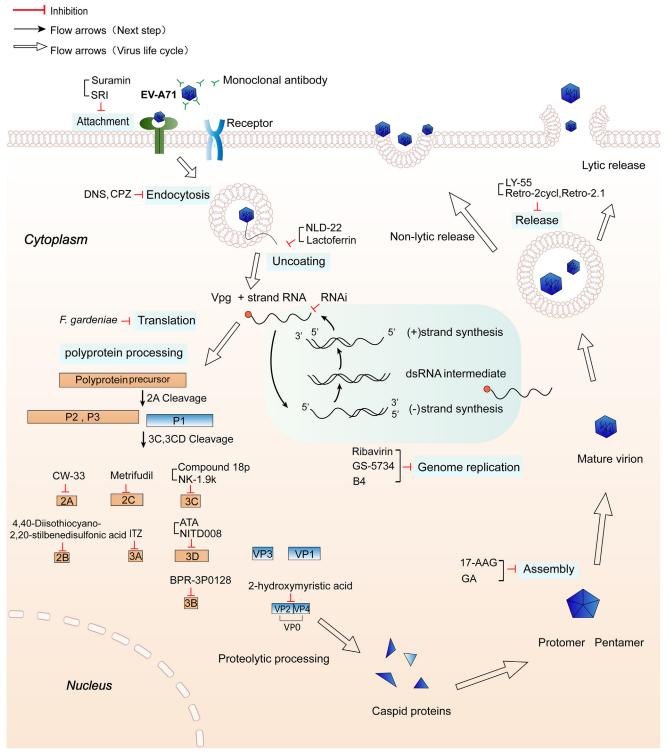
covalently links to the EV-A71 genome. The parent virus RNA is a messenger RNA and it is translated into a single large polyprotein. Next, the viral protease 2A processes the polyprotein into the following 3 parts: 1 structural (P1) and 2 nonstructural (P2 and P3) regions. Among these 3 regions, P1 is further proteolyzed into VP0 (VP2 and VP4), VP1, and VP3. P2 and P3 are ultimately converted into 7 replicate proteins (P2-2A, -2B, and -2C; P3-3A, -3B, -3C, and -3D) by viral proteinases 3C and 3CD.<sup>15</sup>

The virus uses the error-prone RNA-dependent RNA polymerase 3D to replicate the viral genome in a vesicular membrane structure. The virus uses the negative-strand (–) RNA as a template to synthesize new positive-strand (+) RNA. The newly generated positive-strand (+) RNA enters the following steps of the viral life cycle: transcription, translation, and replication. Then, the newly synthesized genomic RNA binds to the viral capsid proteins to form new virions—virus maturation heralds when VP0 is genome inducible for cleavage into VP4 and VP2. Finally, mature viral particles are released extracellularly through cellular lytic or nonlytic pathways.<sup>16</sup>

# Pathogenesis and Pathophysiology of Enterovirus A71 Infection

#### Virus Infection Path

The mechanism of EV-A71 pathogenesis has been studied extensively. Humans are the only host and source of infection for EV-A71. EV-A71 infects an organism and initiates the viral replication cycle after successfully binding to specific receptors on the host, such as gastrointestinal, respiratory, and dendritic cells. EV-A71 initially replicates in the lymphoid tissue of the oropharyngeal cavity and small intestine. Further, it proliferates in the deep cervical and mesenteric nodes before entering the reticuloendothelial system, heart, lungs, skin, mucous membranes, and central



# Figure 3.

An overview of the enterovirus life cycle and inhibitors targeting EV-A71 infection. The enterovirus life cycle comprises virus entry (attachment, endocytosis, and uncoating), translation, polyprotein processing, replication, and assembly and release of new virions. If any of these stages is inhibited, the subsequent stages are affected. Representative inhibitors of EV71 are shown in the figure. ATA, aurintricarboxylic acid; CPZ, chlorpromazine; DNS, dynasore; dsRNA, double-stranded RNA; EV-A71, enterovirus A71; GS-5734, remdesivir; SRI, sophoridine; RNAi, RNA interference.

nervous system (CNS). EV-A71 can also be transmitted along the nerve and into the CNS via retrograde axonal transport.

EV-A71 can enter the CNS through 2 routes. The virus can either enter the CNS by the blood–brain barrier or by peripheral nerves through retrograde axonal transport.<sup>17</sup> The EV-A71 virus also transmits to the CNS by peripheral motor nerves, and the skeletal muscle gets immediately infected by the CNS not only by motor neurons but also by other neural pathways.

#### Factors Related to Viral Infection

The infection of EV-A71 depends on multiple effects of the virus, host, and environment. One of the factors affecting EV-A71 infection is the virulence of the virus. The severity of clinical symptoms caused by EV-A71 varies between genotypes. The change in variation in the sequences in RNA causes some neurologic infections. Li et al<sup>18</sup> suggested that an L97R alteration in the VP1 protein increases the neuronal tropism of EV71, although the alterations in VP1, the 5' noncoding region (NCR), and protease 2A affect viral virulence.<sup>18,19</sup> Meanwhile, coinfection with EV-A71 and other viruses, such as dengue encephalitis, can affect the severity of viral infections.

Additionally, host susceptibility also influences the virus infection.<sup>20</sup> Some anatomical data show that viral replication in virus-infected patients does not coincide with the site of lesion onset, suggesting that there may be present other pathogenic mechanisms, such as host immunity.<sup>21</sup> One genetic study in Taiwan reported that human leukocyte antigen-A33 is associated with increased susceptibility to EV-A71 infection.<sup>22</sup> It has also been reported that abnormal cytokine activation produces severe inflammation in the lungs.<sup>2</sup> From several studies, it was found that children with severe EV-A71 encephalitis have a cytotoxic T-lymphocyte antigen haplotype (cytotoxic T-lymphocyte–associated protein 4).<sup>23</sup> Currently, tissue-specific viral virulence is poorly understood in cell-based systems and animal models; it needs more study in the future.

In particular, genes involved in mediating EV-A71 virus escape from host intrinsic or adaptive immune response monitoring are closely related to EV-A71 susceptibility.<sup>24</sup> For example, interleukin (IL)-6 and monocyte chemoattractant protein 1 genes may affect the risk and severity of EV-A71 infection by affecting their gene expression and regulating inflammatory response. Most importantly, the level of cytokines fluctuates a lot between healthy volunteers, mild cases, and severe EV-A71-infected HFMD patients with complications, which indicates that cytokines may play a critical role in the progress of EV-A71 infection. Recently, several cytokines or chemokines such as tumor necrosis factor (TNF)- $\alpha$ , interferon (IFN)- $\gamma$ , multiple ILs , monocyte chemoattractant protein 1, granulocyte colony-stimulating factor, and HMGB1 have been reported to be associated with severe EV-A71 infection.<sup>25</sup>

# Pathological Mechanisms of Enterovirus A71 Causing Severe Central Nervous System Diseases

EV-A71 occasionally involves the CNS and induces diverse neurologic complications, such as brainstem encephalitis, aseptic meningitis, and acute flaccid paralysis. Among those complications, brainstem encephalitis is the most critical neurologic manifestation because it can cause neurogenic pulmonary hemorrhage/edema leading to death. Despite the discovery of receptors for EV-A71 in human cells, such as the scavenger receptor B2 and P-selection glycoprotein ligand 1, it is not known why EV-A71 infection predominantly involves the brainstem.<sup>26</sup>

Cytokines, as a part of innate immunity, favor the development of antiviral and type I helper T-lymphocyte immune responses. Cytokines and chemokines play an important role in the pathogenesis of EV-A71 brain stem encephalitis. Both the CNS and the systemic inflammatory responses to infection play important, but distinctly different, roles in the pathogenesis of EV-A71 pulmonary edema.<sup>27</sup>

Endothelin 1, a potent vasoconstrictor, can induce pulmonary edema in rats via intrathecal injections. Endothelin 1 in the CNS may play a role in the development of neurogenic pulmonary edema in children with EV-A71 infection and could be used as a biomarker or therapeutic target for neurogenic pulmonary edema in EV-A71 encephalitis.<sup>28</sup>

However, the mechanisms through which EV-A71 causes neurologic diseases have not been fully explored. Accumulated evidence indicates that EV-A71 infection triggers a plethora of interactive signaling pathways, resulting in host immune evasion and inflammatory response. EV-A71 can activate cellular signaling networks, including multiple cell surface and intracellular receptors, intracellular kinases, calcium flux, and transcription factors that regulate antiviral innate immunity and inflammatory response. Cellular signaling plays a critical role in the regulation of host innate immune and inflammatory pathogenesis.

#### Histopathologic Characteristics of Enterovirus A71 Infection

Most EV-A71 infections will resolve spontaneously, with a small proportion progressing to severe HFMD. Patients with severe HFMD have inflammatory lesions in several body organs, with the brainstem considered the most vulnerable area. The brainstem and spinal cord are the main target areas in pathological CNS lesions caused by EV-A71.<sup>29</sup> Autopsy and magnetic resonance imaging of patients with EV-A71 brainstem encephalitis showed a widespread inflammatory response in the gray matter of the spinal cord and throughout the medulla oblongata. The pathology suggests extensive vascular dilatation and bruising on the brain's surface, with a high degree of edema in the brain tissue and numerous foci of softening centered on the brainstem and medulla oblongata, satellite phenomena of nerve cells, and the phenomenon of neurophilic cells.<sup>21,27,30,31</sup> Immunohistochemistry staining showed that inflammatory cells in the CNS mainly were CD68+ macrophages/microglia or CD15+ neutrophils, widely distributed in areas with extreme inflammatory changes. EV-A71 may spread from the site of primary infection to the CNS via the inflammatory cell migration pathway.<sup>32</sup>

Neurogenic pulmonary edema is a significant complication of EV-A71 infection and a cause of death. This complication is often secondary to various CNS injuries, such as encephalitis. Pathological analysis showed dilated and congested capillaries in the alveolar walls, widened alveolar septa, massive inflammatory cell infiltration in the interstitium, massive serum exudation in the alveolar cavity, hyaline membrane formation, compensatory emphysematous changes in some lung tissues, and reactive hyperplasia of lymph nodes next to the hilar bronchi. Some investigators found that IL-1B, -6, -10, and -13, TNF- $\alpha$ , and IFN- $\gamma$  were significantly increased in patients with EV-A71 brainstem encephalitis accompanied by pulmonary edema.<sup>7,20,27</sup>

Pathological observation of EV-A71 infection also reveals a heart with marked focal cardiomyocyte edema and an infiltrate of scattered lymphocytes, a few monocytes, and neutrophils in the intercellular plasm. The spleen showed massive parenchymal cell necrosis or reactive hyperplasia of lymph nodes. In addition, there was reactive hyperplasia of mesenteric lymph nodes, enlarged lymphoid follicles, and histiocytosis.<sup>21</sup> Although no significant pathological changes were seen in the digestive system and heart in most studies, enlarged tonsils , mesenteric lymph nodes, and varying degrees of inflammatory swelling in the heart were present in some deaths. Based on the pathogenesis and pathological data of EV-A71 infection, we recommend that the treatment of EV-A71 infection observes the principles of early prevention, early recognition, and early treatment, paying attention to its clinical symptoms, improving auxiliary examinations, and the rational use of drugs based on the protection of the body's immune system, thus preventing and treating the CNS and other complications. Additionally, a study found a strong signal of EV-A71 RNA in the epithelial cells of renal tubules.<sup>32,33</sup>

These mechanisms of viral infections and their pathology lay the foundation for their prevention and treatment in the clinical setting.

# **Antiviral Agents for Enterovirus A71 Infection**

The ability of a virus to successfully infect cells depends on its ability to enter and go through a life cycle process (entry, translation, polyprotein processing, replication, assembly, and release of new virions). It means that several steps exist, and each of them represents a target. The key steps of the life cycle provide insights into designing and developing effective antivirals.<sup>34</sup> In the following sections, this review focuses on the inhibitors targeting EV-A71 and highlights examples of each category of inhibitors (Table and Fig. 3).

# **Inhibitors of Virus Entry**

Virus entry is the first step in viral infection, which involves virus attachment, endocytosis, and uncoating. The binding of the virus to the receptor is critical to the tropism of the virus. Known EV-A71 receptors include the human scavenger receptor class B2 (hSCARB2),<sup>35</sup> P-selectin glycoprotein ligand 1 (PSGL-1),<sup>36</sup> heparan sulfate,<sup>37</sup> sialylated glycan,<sup>38</sup> vimentin,<sup>39</sup> nucleolin,<sup>40</sup> annexin II,<sup>41</sup> cyclophilin A (Cyp A),<sup>42</sup> human tryptophanyl aminoacyl-tRNA synthetase,<sup>43</sup> heat shock protein 90 (HSP90),<sup>44</sup> fibronectin,<sup>45</sup> and prohibitin.<sup>46</sup> Some receptors play different roles during viral infection. Among them, hSCARB2, Cyp A, heparan sulfate, sialy-lated glycan, PSGL-1, and human tryptophanyl aminoacyl-tRNA synthetase are related to virus uncoating. Targeting EV-A71 entry is a promising strategy to inhibit viral infection. Currently, there are many antivirals targeting virus entry.

#### Virus Attachment

Attachment is the first step of virus entry. The virus binds to specific receptors on the surface of the host, which mediates the process of virus attachment. Blocking adsorption is a very effective method to inhibit viral infection. Some drugs targeting virus attachment are described below.

Wang et al<sup>47</sup> showed that suramin inhibited the early infection of EV-A71 by blocking viral adsorption. Suramin has been used in clinical practice for decades, and its safety is not a concern. Suramin is currently in phase I of the HMFD clinical trial study and it is worth digging into its antiviral activity.<sup>48,49</sup> NF449, a suramin analog, exhibits excellent anti–EV-A71 activity.<sup>50</sup> LJ04 and melittin show outstanding virucidal activity against EV-A71.<sup>51,52</sup> Melittin is currently in phase II of clinical trials for osteochondrosis.<sup>53</sup> Arg-Gly-Asp-Ser can bind to fibronectin and block viral entry. Additionally,

SP40 and L-SP40, peptides designed after the VP1 protein of EV-A71, can inhibit viral attachment.<sup>54</sup> Sophoridine and brilliant black BN (E151) prevent EV-A71 infection by inhibiting viral adsorption.<sup>55,56</sup> However, no clinical trial information is available for drugs other than suramin and melittin, and their safety needs further investigation.

#### Endocytosis and Virus Uncoating

Enteroviruses enter cells via crossing lipid membrane barriers through endocytosis. Then, the virus binds to specific receptors or induces a change in the pH of the cell, thereby helping it to uncoat.<sup>13</sup> Some drugs have been found to inhibit viral entry by targeting endocytosis and uncoating. Endocytosis inhibitors, such as chlor-promazine and dynasore, can inhibit EV-A71 infection.<sup>57</sup> The bovine and human lactoferrin, rosmarinic acid, and NLD-22 protect susceptible hosts by preventing viral uncoating.<sup>58-62</sup> Additionally, compound 11, an inhibit or of Cyp A, can interact with the H-I loop of VP1 to regulate viral uncoating.<sup>42,63</sup> Vapendavir, a novel enteroviral capsid binder, has broad antiviral activity.<sup>64</sup>

VP1, the leading antigen-binding site of EV-A71, is relatively conservative; the development of capsid-binding inhibitors can block the interaction between EV-A71 and the host and improve the drug resistance of EV-A71 mutations.<sup>65</sup> For example, compound 11 may be one of the more clinically promising target compounds for the subsequent development of viral entry inhibitors. Additionally, researchers can enlarge the study of viral infection receptors, which can provide ideas for designing and developing antivirals.

#### **Inhibitors of Viral Translation**

When the virus genome enters the cytoplasm, the virus starts the translation phase of its life cycle. The internal ribosome entry site (IRES) is involved in the translation process of the genome. When the translation process is blocked, the viral replication is interrupted. Therefore, the translation stage of the virus is also one of the targets of antiviral design.<sup>66</sup>

Magnesium lithospermate B (MLB) is the main component of Danshen pill injection. Danshen pill injection, a traditional Chinese medicine, was approved in May 2005. MLB can inhibit viral IRES.<sup>67</sup> Quinacrine interrupts RNA transcription and viral protein translation by intercalating into viral nucleic acids.<sup>68</sup> Kaempferol has an anti–EV-A71 effect, and its antiviral mechanism occurs by inhibiting viral translation.<sup>69</sup>

From the point of view of medical application, efficacious inhibitors of viral translation can not only specifically inhibit IRES but also affect the activity of the eukaryotic promoter. As with MLB, it exerts antiviral activity without affecting the action of the eukaryotic promoter in the host. In China, no severe adverse drug reactions have occurred in Danshen pill injection in the past 8 years, so MLB has greater clinical potential among the 3 viral translation inhibitors.<sup>70,71</sup>

# **Inhibitors of Viral Polyprotein Processing**

### Enterovirus A71 Structural Proteins

The role of viral structural proteins is mainly to assemble into the viral capsid structure, also known as the capsid protein. Viral

#### Table

6

The detailed list and classification of EV-A71 inhibitors: classes, effectivity, test in cell lines and animal models, and clinical trials

No.	Compound	Target	<sup>a</sup> EC <sub>50</sub> / <sup>b</sup> IC <sub>50</sub>	Cell line/animal	<sup>c</sup> Clinical trials
Inhibitors of entry					
1	Suramin	Attachment	40 µM	RD	Phase I (HFMD)
2	NF449	Attachment	6.7 μM	RD	-
3	Melittin	Directly inactivates EV-A71	0.76 μg/mL	HeLa	Phase II (tumor-induced osteomalacia)
4	SP40 and L-SP40	Attachment	6-9.3 mM	RD, HeLa, HT-29	-
5	LJ04	Directly inactivates EV-A71	$24.3 \pm 2.4 \ \mu M$	MA104	-
6	RGDS	Fibronectin	5 mg/mL, 5 mg/kg	RD, Suckling mice	-
7	SRI	Attachment	31.25 μg/mL	Vero	-
8	E151	VP1	2.39-28.12 μM	RD	-
9	CPZ, DNS	Endocytosis	20 μM, 80μM	HepG2, Vero, RD	Phase III (COVID-19)
10	Lactoferrin	Heparin sulfate glycosaminoglycans	34.5 g/mL	SK-N-SH, RD	Phase II (HIV)
Phase 2 (SARS-CoV-2)					
11	RA	VP1	4.33 ± 0.18 μM	RD	Phase IV (osteoarthritis of the knee)
12	NLD-22	Uncoating	25 pM	RD	-
13	Compound 11	Сур А	0.37 ± 0.17 μM	Spleen cells	-
14	Vapendavir	Capsid protein	0.7 μM	RD	Phase I (healthy)
Phase 2 (asthma)					
nhibitors of viral translatio	on				
15	MLB	IRES	0.09 mM	RD	Phase IV (angina)
16	Quinacrine	RNA transcription	9.71 mM	RD	Phase II (prostatic cancer)
17	Kaempferol	IRES	35 μM	RD	Phase I (healthy)
Inhibitors of viral polyprote	ein				
processing	LUCTN .	24	0.C. M	11-1 -	
18	LVLQTM	2A	9.6 μM	HeLa	-
19	CW-33	2A	171.2 μM	RD	-
20	4,40-Diisothiocyano-2,20- stilbenedisulfonic acid	28	1.76 μg/mL	RD	-
21	Adenosine analogs: metridil				
N6-benzyladenosine	2C	1.30 μM			
0.10 mM	RD	-			
22	AN-12-H5	3A	0.55 μΜ	DLD-1 intestinal	-
23	GW5074	3A	2.00 μM	DLD-1	Phase I (solid tumor)
24	ITZ	3A	1.15 μΜ	RD	Phase I (RSV/chronic hepatitis B)
25	BPR-3P0128	3B	0.0029 μM	RD	-
26	AG7088	3C	0.014 μM	RD	-
27	Compound 18p	3C	$0.030 \pm 0.02 \ \mu M$	RD, Vero	-
28	SG85, PI SG85	3C	180 nM		
0.039-0.200 μM	RD	-			
29	NK-1.8k, NK-1.9k	3C	34.5 nM, 37.00 nM	RD, Vero	-
30	Quercetin	3C	12.1 μM	RD, Vero	Phase III (COVID-19)
Phase 1 (chronic hepatitis (					
31	NITD008, ppp-NITD008	3D	0.625 μΜ	RD	-
32	ATA	3D	2.9 μM	RD, Vero	-
33	DTriP-22	3D	0.30 μΜ	RD, Vero, HeLa	-
34	GPC-N114	3D	0.1-1 μM	RD, Vero	-

Inhibitors of virus replica	ation				
35	Ribavirin	RNA	65 μg/mL	RD	Approved for marketing (broad-spectrum
					antiviral drugs)
36	Qramine derivatives 4s	Early viral replication	9.1 μg/mL	RD, Vero	-
37	Chebulagic acid	Genome replication	12.5 μg/mL	RD	-
38	Apigenin	Viral RNA transacting factors	10.3 mM	RD, Vero	-
39	GS-5734	Genome replication	0.991 μM	HeLa	Approved for marketing (SARS-CoV2)
Phase 2 (Ebola/HIV)					
40	B4	IFN-β	$24.95 \pm 0.05 \ \mu M$	RD, Vero, ICR mice	-
41	Aloe-emodin	ISGFs	0.14-0.52 μg/mL	HL-CZ, TE-671	-
42	CP	I IFN	$0.39 \pm 0.07 \ \mu g/mL$	HeLa	-
43	Polyriboinosinic acid (poly(I : C)	IFNs	65 nM	RD	Early phase I (breast cancer)
Phase I (recurrent gliobla	astoma)				
44	Xiyanping	IFNs	5-10 mg/kg	ICR mice	Approved for marketing (anti-inflammatory viral)
45	Minocycline	IL6, G-CSF	100-300 μg/mL	RD, U-87MG, THP-1	Approved for marketing (antibiotics)
46	Paris polyphylla Smith	IL-6	78.46 ± 2.80 g/mL	RD	-
47	PD169316	p38	20 μM	RD, HeLa	-
48	Formononetin	ERK, p38, and JNK	$3.98 \pm 0.80 \ \mu M$	SK-N-SH	-
49	Resveratrol	ΙΚΚα, ΙΚΚβ, ΝΓ-κΒ p65	20.2 μΜ	RD	Approved for marketing (anti-inflammatory cancer)
50	Corydaline	JNK, P38, MAPK	25.23 ± 6.60 μM	Vero	-
51	Compound 18	MEK1	10 μM	RD, HEK293	-
52	Berberine	AKT, PI3KIII, MEK/ERK	7.43-10.25 μM	Vero	Phase IV (COVID-19)
Phase III (cirrhosis due to hepatitis B)	0				
53	GS-9620	PI3K-AKT			
NF-ĸB	7.43-10.25 μM				
6.0 mg/kg	Vero				
ICR mice	Phase 1 (hepatitis B/C)				
Phase 2 (HIV)					
54	SeNPs@OT	Mitochondrial pathway	9.8-µM SeNPs and 20-nM oseltamivir	U251	-
55	EGCG and GCG	ROS	10 μM	Vero	Phase I (COVID-19)
56	Luteolin	ROS	Approximately 10 µM	Vero, RD	Early phase I (tongue neoplasms carcinoma
57	Gallic acid	ROS	0.76 μg/mL	Vero	-
58	PDTC	UPS	25-50 μΜ	Vero	-
Inhibitors of virion assen release	nbly and				
59	Retro-2cycl and Retro-2.1	Viral release	12.56 mM and 0.05 mM	293S	-
60	2-Hydroxymyristic acid	VP0	50 μM	RD	-
61	LY-55	Autophagy	$2.22 \pm 0.44 \ \mu M$	Vero	-
62	Anti-FLIP peptide	cFLIP	0-10 μg/mL	MRC5	-
63	GA	ΗSP90β	2-20 μM	RD, Vero	-
64	17-AAG	ΗSP90β	0.5-0.2 μg/kg	RD, Vero, hSCARB2- transgenic mice	Phase II (kidney cancer)
Other EV-A71 inhibitors					
65	Raoulic acid	Replication	<0.1 µg/mL	Vero	-
66	Ursolic acid	Replication	0.5 μg/mL	BCC-1/KMC	Phase III (sarcopenia)
67	Glycyrrhizic acid	Event(s) post virus cell entry	1.01	Vero	Phase IV (immune thrombocytopenia)
68	Corilagin		5.6 μg/mL	Vero	-

7

(continued on next page)

ontinue	
9	
el el	
5	

( p

No.	Compound	Target	$^{a}EC_{50}/^{b}IC_{50}$	Cell line/animal	<sup>c</sup> Clinical trials
69	Honeysuckle	<i>let-7a</i> and viral genome	100-600 nM	RD, SK-N-SH	Phase III (irritable bowel syndrome)
70	mAbs (3A12, 2A10)	3D	1	Vero, BALB/c ICR mice	
71	D5, H7, C4	VP1 GH loop	0.203, 0.287, 0.952 µg/mL	RD, Vero, Jurkat T	1
72	mAb51	VP1	10 µg/g	RD, BALB/c mice	1
73	mAbs 22 and 24	VP1	1	RD, BALB/c mice	1
74	JL2	SCARB2	2 μg/mL	293-hSCARB2 cells	1
75	si-2C, si-3C, si-3D	2C, 3C, 3Dpol	I	RD	1
76	si-3' UTR	3' UTR	I	RD	1
77	si-5' UTR	5' UTR	1	RD	1
78	siRNA-69, siRNA-294, siRNA-319	2A	I	RD	1
79	miR-9	NF-kB	1	RD, Vero, HT-29	
ICR mice	I				

17-AAG, 17-allyamino-17-demethoxygeldanamycin; AG7088, rupintrivir; AKT, protein kinase B; ATA, aurintricarboxylic acid; COVID-19, coronavirus disease 2019; CP, Cortex phellodendri; CPZ, chlorpromazine; Cyp A, cyclophilin A; DNS, dynasore; EGCG, epigallocatechin gallate; ERK, extracellular signal-regulated kinase; EV–A71, enterovirus A71; GA, geldanamycin; GCG, gallocatechin gallate; G-CSF, granulocyte colony-stimulating factor; GS-5734, remdesivir; HFMD, hand, foot, and mouth disease; HIV, human immunodeficiency virus; hSCARB2, human scavenger receptor class B2; HSP90, heat shock protein 90; IFN, interferon; IL, interleukin; IKK, inhibitor of kappa kinase; IRES, internal ribosome entry site; ISFG, IFN-stimulated gene factor; ITZ, itraconazole; JNK, c-jun amino-terminal kinase; mAb, monoclonal antibody; MEK, MAP/ERK kinase; miR, microRNA; MLB, magnesium lithospermate B; NF+KB, nuclear factor kappa B; PI3K, phosphatidylinositol 3-kinase; PI3KIII, phosphatidylinositol 3-kinase, class III; PTDC, pyrrolidine dithiocarbamate; RA, rosmarinic acid; RGDS, Arg-Gly-Asp-Ser; RNAi, interference; ROS, reactive oxygen species; RSV, respiratory syncytial virus; SARS-CoV-2, severe acute respiratory syndrome-related coronavirus 2; SeNPs, selenium nanoparticles; siRNA, small interfering RNA; SRI sophoridine; UPS, ubiguitin-proteasome system; UTR, untranslated region Concentration for 50% of maximal effect.

Half-maximal inhibitory concentration.

р

Search website: https://clinicaltrials.gov/ct2/home

antigenic diversity is affected by the structural changes of the capsid proteins VP1 to VP3. Capsid proteins link to viral entry, assembly, and release. Therefore, inhibitors of capsid proteins are classified as inhibitors of viral entry, assembly, and release. It is not explained in detail in this study.

# Enterovirus A71 Nonstructural Proteins

# 2A

The 2A protease has cysteine protease activity and it hydrolyzes the EV-A71 RNA-translated polyprotein into the following 3 consecutive parts: P1, P2, and P3. 2Apro also regulates EV-A71 replication by cleaving 3C and 3D proteins and eukaryotic initiation factor 4GI. Given the properties of EV-A71 2Apro, it has the potential to be an antiviral target.<sup>72</sup> LVLQTM, a pseudosubstrate of 2Apro, can bind to the substrate-binding pocket of EV-A71, interfering with the replication process of EV-A71.73,74 CW-33 specifically binds to the active site of the viral 2A protease, thereby inhibiting EV-A71 replication.<sup>75</sup>

# 2B

The 2B protein has viral porin activity and promotes viral transmembrane and genome replication. The 2B protein can also interact with proapoptotic proteins to induce apoptosis and release viral particles. The function of the 2B protein in the virus provides ideas for finding anti-EV-A71 drug targets. Xue et al<sup>76</sup> indicated that EV-A71 2Bpro might mediate chloride-dependent currents in RD cells. 4,40-Diisothiocyano-2,20-stilbenedisulfonic acid can effectively inhibit chloride-dependent currents, thereby preventing EV-A71 infection.

# 2C

The EV-A71 2C protein is closely related to many critical steps in the viral life cycle, such as viral uncoating and rearrangement of cell membranes. EV-A71 2Cpro also inhibits the phosphorylation of inhibitor of kappa B kinase  $\beta$  (IKK $\beta$ ), interfering with the nuclear factor kappa B (NF-κB) signaling pathway.<sup>77</sup> N-(2-methylphenyl)methyladenosine and N6-benzyladenosine effectively reduce the probability of host infection with the virus by interacting with 2C.<sup>13,5</sup>

# 3A

The EV-A71 3A protein is a membrane-bound protein. It can interfere with endoplasmic reticulum division, hinder membrane protein transport function, and promote viral RNA synthesis.<sup>78</sup> Additionally, 3Apro can inhibit the expression of cytokines, such as IL-6 and IL-8, and inhibit the host's antiviral immune function. AN-12-H5 interferes with the early stages of viral infection by targeting EV-A71 3Apro.<sup>79</sup> Itraconazole (ITZ) and GW5074 inhibit the synthesis of EV-A71 new particles by inhibiting 3A. ITZ is currently in phase I of clinical trials of respiratory syncytial virus (RSV). It has greater clinical potential for anti-EV71 development than the other 3A inhibitors.<sup>50,7</sup>

# 3B

The EV-A71 3B protein is also known as the VPg protein. The conserved tyrosine residue at position 3 of the VPg protein forms a phosphodiester bond with the uridine monophosphate at the end of the genome, which can link to the 5' end of the viral RNA. Then, 3D polymerase-catalyzed VPg uridylation is used as a primer to promote RNA synthesis. Currently, there are relatively few studies on agents targeting 3B.<sup>81</sup> BPR-3P0128, an inhibitor of multiple molecular targeting sites, can inhibit EV-A71 proliferation by targeting EV-A71 VPg uridylation and RNA-dependent RNA polymerase.82

#### 3C

The EV-A71 3C protein has multiple functions, such as 3Cpro catalyzes the cleavage of viral precursor proteins and promotes cell apoptosis.<sup>13</sup> 3Cpro can interfere with the polyadenylation of cell RNA by regulating *CstF-64*, thereby enhancing viral infection.<sup>83</sup> Owing to the critical role of 3Cpro in viral replication, it has become a popular target for antiviral research.

Rupintrivir and compound 18p target 3Cpro of EV-A71. The latter has good pharmacokinetic properties in the body.<sup>84,85</sup> Peptidyl aldehyde NK-1.8k and a highly specific  $\alpha$ -hydroxynitrile derivative NK-1.9k suppress EV-A71 infection by targeting protease 3C. Compared with NK-1.8k, NK-1.9k has more obvious antiviral properties.<sup>86,87</sup> Additionally, SG85, PI SG85, and quercetin can also inhibit the activity of EV-A71 3Cpro. Among the 3C inhibitors, only quercetin has clinical trial stage information. It is currently in phase III of the COVID-19 clinical trial and has the greatest potential for anti-EV-A71 clinical development.<sup>64,88-90</sup>

#### 3D

The 3D protein, a polyproteinase (RNA-dependent RNA polymerase), uridylylates VPg and uses VPg-pUpU as a primer to initiate virus replication and assists in the elongation of viral RNA strands. 3D is considered an attractive target for drug development.<sup>91</sup> NITD008 inhibits the proliferation of enteroviruses. The triphosphorylated product of NITD008, ppp-NITD008, inhibits EV-A71 infection by inhibiting viral RNA-dependent RNA polymerase activity.<sup>92</sup> Aurintricarboxylic acid and GPC-N114 exhibited inhibitory activity against EV-A71 by interfering with the viral 3D polymerase.<sup>16,93,94</sup> DTriP-22, a novel nonnucleoside analog targeting EV-A71 3Dpol, inhibited EV-A71 replication by reducing the accumulation of RNA.<sup>95</sup> However, the 3D inhibitors have not yet entered clinical studies. There is much research on inhibitors targeting viral proteins, and most drugs focus on 3Cpro. Among them, AG7088 and quercetin have entered clinical trials. For example, a pharmacokinetic-pharmacodynamic study of quercetin in healthy adults and patients with hypercoagulable states has been completed.<sup>96</sup> The advantages of inhibitors targeting viral proteins are more specific and less toxic, but the viral spectrum is narrow, and the risk of developing resistance is high. Combining antivirals with different mechanisms of action not only improves drug efficacy but also reduces the occurrence of resistance. Additionally, most inhibitors of viral polyprotein processing studies have been limited to the cellular level and have yet to be clinically validated. Therefore, investigators can prioritize compounds that have already entered clinical trials as targets for further studies.<sup>97,98</sup>

## **Inhibitors of Virus Replication**

This section introduces the targets of existing anti-EV-A71 replication, including IFNs; ILs; mitogen-activated protein kinase (MAPK), phosphatidylinositol 3-kinase (PI3K), and protein kinase B (Akt) signaling pathways; the oxidative stress pathway; the ubiquitin-proteasome system; and so on. Furthermore, it focuses on the inhibitors targeting EV-A71 replication and highlights examples of each category of inhibitors (Fig. 4).

### Virus Genome Replication

The core enzyme of genome replication is the viral RNAdependent RNA polymerase 3Dpol, the replication primer is the viral peptide 3B coupled to 2 uridines, and the viral replication properties are highly conserved. Genome replication is a promising target for researching and designing broad-spectrum antiviral agents.<sup>99</sup>

Ribavirin is an RNA viral mutagen. It exerts broad-spectrum antiviral activity by inducing lethal mutations in the viral genetic material. Ribavirin is one of the most commonly used broadspectrum antivirals in clinical practice and is susceptible to drug resistance. Currently, the results of clinical trials of phase IV of ribavirin plus pediatric oral solution on HMFD is complete.<sup>100-102</sup> Gramine derivatives 4s and chebulagic acid protect the host by inhibiting RNA replication.<sup>103,104</sup> Apigenin protects the host by disrupting the binding of viral RNA to host transacting factors.<sup>105</sup> Additionally, Ye et al<sup>106</sup> showed that remdesivir (GS-5734)-hindered RNA replication exhibited vigorous antiviral activity against EV-A71. GS-5734 has been approved for the treatment of SARS-CoV2 infections with a favorable safety profile and has high potential for clinical development among these drugs that inhibit EV-A71 genome replication.

# Interferons

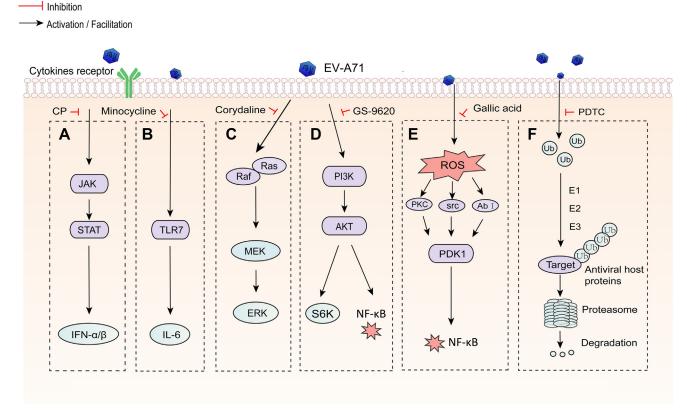
IFNs are divided into  $\alpha$  type,  $\beta$  type, and  $\gamma$  type. IFNs play an essential role in the antiviral defense of cells. IFNs can protect EV-A71-infected hosts by modulating the host's innate immune function and interrupting viral replication.<sup>107,108</sup> Anemoside B4 and aloe-emodin play an antiviral role by upregulating the expression of IFN- $\beta$ .<sup>109,110</sup> Cortex phellodendri aqueous extract has broad-spectrum antiviral effects by inducing the production of IFNs.<sup>111</sup> Additionally, polyriboinosinic acid acts as a potent IFN inducer that induces the production of type I IFN.<sup>107</sup> Andrographolide sulfonate (Trade name: Xiyanping injection) has inactivating effects on the influenza virus and EV-A71. It can regulate the immune system of host by regulating the secretion of inflammatory factors. Currently, Xiyanping injection is used clinically for antipyretic, anti-inflammatory, and antiviral treatment.<sup>112</sup> Additionally, IFN- $\alpha$  spray or nebulization is effective in the early stages of treating EV-A71 infection.<sup>1,25</sup>

#### Interleukin

IL is a cytokine secreted by a variety of cells. IL is involved in various physiological and pathological body responses, such as the role of transmitting information and the activation and regulation of the immune system. When a virus infects the body, the host protects itself by producing IL to fight virus invasion.<sup>25</sup> Minocycline reduced EV-A71 offspring production by upregulating IL-6 and granulocyte colony-stimulating factor levels. Minocycline also alleviates CNS complications by reducing TNF levels in the host CNS. Minocycline is approved for marketing as an antibiotic with a proven safety profile. Researchers can continue to study it as an anti-EV-A71 clinical candidate.<sup>113</sup> Additionally, Wang et al<sup>114</sup> found that Paris polyphylla Smith antagonizes EV-A71 replication mainly through the regulation of IL-6 levels.

#### Mitogen-Activated Protein Kinase

After a virus infects a cell, the toll-like receptor changes its conformation and activates a series of signal transduction, especially MAPK activation. Then, the signal pathways mediated by extracellular signal-regulated kinase, c-jun amino-terminal kinase (JNK), and p38 are activated to start the transcription of various inflammatory factors. MAPK is one of the most studied pathways associated with EV-A71 entry and replication.<sup>115</sup>



#### Figure 4.

An overview of the signal pathways and inhibitors targeting EV-A71 infection. Representative inhibitors of EV-A71 are shown in the figure. (A) The interferon pathway. (B) The interleukin pathway. (C) The MAPK pathway. (D) The PI3K/Akt pathway. (E) The oxidative stress pathway. (F) The ubiquitin-proteasome system. Abl, ableson protein tyrosine kinase; Akt, protein kinase B; dsRNA, double-stranded RNA; E1, ubiquitin-activating enzyme; E2, ubiquitin-conjugating enzyme; E3, ubiquitin ligase; ERK, extracellular regulated kinase; EV-A71, enterovirus A71; IFN- $\alpha/\beta$ , interferon alpha and interferon beta; IL-6, interleukin 6; JAK, janus kinase; MAPK, mitogen-activated protein kinase; MEK, MAP/ERK kinase; NF- $\kappa$ B, nuclear factor kappa B; PDK1, 3-phosphoinositide-dependent kinase 1; PI3K, phosphatidylinositol 3-kinase; PKC, protein kinase C; ROS, reactive oxygen species; S6K, S6 kinase; src, steroid receptor coactivators; STAT, signal transducer and activator of transcription; TLR7, toll-like receptor 7; Ub, ubiquitin.

PD169316, a p38 inhibitor, inhibits cell apoptosis and reduces the release of EV-A71.<sup>116</sup> Formononetin, corydaline, and compound 18 interfere with EV-A71 replication and limit virus release by inhibiting the activation of extracellular signal-regulated kinase, p38, and JNK signaling pathways.<sup>117-119</sup> Resveratrol inhibits the proliferation of EV-A71 by inhibiting the phosphorylation of IKKα, IKKβ, and NF-κB p65 and preventing the secretion of antiviral cytokines. Resveratrol is approved for its anti-inflammatory/ anticancer properties and an excellent safety profile and has the potential to be developed into a clinical anti-EV71 drug.<sup>120</sup>

#### PI3K/Akt

PI3K/Akt is a canonical signaling pathway associated with phosphatidylinositol and plays an essential role in promoting cell survival. This pathway is activated when EV-A71 stimulates cells.<sup>121</sup> Wang et al<sup>122</sup> found that berberine inhibited the phosphorylation of AKT, JNK, and PI3KIII. Later, they also showed that berberine derivatives exhibited significant anti–EV-A71 activity.<sup>123</sup> The heterocyclic compound GS-9620 is classified as a selective TLR7 agonist with potent and low toxicity. It inhibits the dissemination of EV-A71 mainly through NF-κB and PI3K/Akt pathways.<sup>124,125</sup> Furthermore, berberine anti–COVID-19 studies are in phase IV of clinical trials and its safety has been

demonstrated. Researchers can prioritize berberine as an anti-EV71 candidate over other PI3K/Akt inhibitors.

#### Reactive Oxygen Species

Reactive oxygen species (ROS) plays a vital role in host selfdefense. ROS is a double-edged sword. When viruses infect cells, they typically turn on their defense mechanisms, producing ROS. When many ROS damage the antioxidant defense system, it will result in the body's oxidative stress response. In some cases, ROS can aid in viral infection.<sup>126</sup>

SeNPs@OT, which are selenium nanoparticles loaded with oseltamivir, reduce EV-A71–induced apoptosis and autophagy of cells through the mitochondrial pathway.<sup>127</sup> Gallic acid, epi-gallocatechin gallate, and gallocatechin gallate inhibit EV-A71 replication in a concentration-dependent manner. Their antiviral effects are closely related to ROS.<sup>128,129</sup> Luteolin prevents EV-A71 infection by inhibiting the generation of intracellular ROS and delaying the apoptosis of cells.<sup>130,131</sup>

# Ubiquitin-Proteasome System

The ubiquitin-proteasome system (UPS) is indispensable in many life activities, such as cell proliferation, differentiation, apoptosis, and immunity. Meanwhile, it is involved in replicating positive-strand RNA viruses, such as EV-A71 and the dengue virus.<sup>132</sup> The UPS is expected to become a new target for the research and development of antivirals. A study by Lin et al<sup>133</sup> showed that pyrrolidine dithiocarbamate could affect EV-A71 transcription, translation, and viral progeny proliferation by downregulating the UPS. However, there is no clinical trial information for pyrrolidine dithiocarbamate. It needed to be further researched if the drug can be further developed into an anti-EV-A71 drug.

Among the inhibitors of virus replication, several drugs are already in clinical trials. The safety of GS-5734 in hospitalized patients with severe COVID-19 has been studied.<sup>134</sup> In addition, minocycline is a broad-spectrum clinical antibiotic, and its security has not needed to be checked.<sup>135</sup> Given their clinical trial results and significant anti-EV-A71 activity, they could be considered for development as clinical antivirals. Additionally, we may consider a prodrug strategy to guide the development of nucleoside antivirals, considering the vital role that nucleoside analogs, such as ribavirin play as antivirals.<sup>136</sup> The target of these inhibitors is mainly the host. They have the advantage of potentially having a broader spectrum of antiviral activity and less risk of developing resistance but have the disadvantage of off-target effects and cytotoxicity.<sup>71</sup> Therefore, we need to increase the potential for the clinical application of compounds through technological innovation, the search for antiviral-related targets, and improved drug screening and design tools.<sup>137</sup>

#### **Inhibitors of Virion Assembly and Release**

The capsid proteins of enteroviruses assemble with newly synthesized genomic RNA to form new infectious mature virions. Some newly generated virions are released extracellularly in vesicles, whereas other virions leave the host via cell apoptosis and lysis. Interestingly, some studies have found that the autophagy of cells can assist the release of viruses in a nonlytic manner.<sup>16,138</sup> When viral assembly and release are blocked, the generation of new viruses is interrupted. Therefore, the viral assembly and release process is a good choice for designing antivirals.

Retro-2cycl and Retro-2.1 block viral release by inhibiting the transport of EV-A71 particles in intracellular vesicles.<sup>139</sup> 2-Hydroxymyristic acid prevents virion maturation by inhibiting the cleavage between VP4 and VP2.<sup>137</sup> The anti-FLIP peptide and lycorine derivative LY-55 prevent the release of EV-A71 progeny by regulating cell autophagy and apoptosis.<sup>140,141</sup> Additionally, geldanamycin, a specific inhibitor of HSP90, reduces the production of EV-A71 progeny by blocking virion assembly. 17-Allyamino-17-demethoxygeldanamycin, an analog of geldanamycin, is consistent with the target of geldanamycin.<sup>134</sup> Data from phase I clinical trials show that 17-allyamino-17-demethoxygeldanamycin can be safely administered in biologically active doses and potentially develop into an anti–EV-A71 drug.<sup>142</sup>

Inhibitors of virion assembly and release belong to the targeted virus drugs. If broad-spectrum antiviral therapy is indiscriminate bombing on the battlefield, then targeted virus therapy is a precise treatment that hits the target. However, the disadvantage of direct-acting viral drugs is prone to drug resistance. Therefore, when screening antivirals, we can pay more attention to medication with a "high-resistance barrier." It is also possible to try to design a fixed-dose combination of anti–EV-A71 drugs that combines different types/targets of anti–EV-A71 into a single drug, which not only prevents the emergence of drug resistance by inhibiting viral replication at multiple points in the viral life cycle and helps to reduce the burden of multiple doses, but also improves patient compliance. For example, fixed-dose combinations of antiretroviral drugs are very successful cases.<sup>71,136</sup>

# **Other Enterovirus A71 Inhibitors**

### Natural Products With Anti-Enterovirus A71 Activity

Many natural products from various plants and animals are widely used in the research against viral infections. Some natural products have shown properties against EV-A71 infection. The advantages of natural products for the treatment of viral infectious diseases are mild action and low cytotoxicity.<sup>143,144</sup> Raoulic acid shows significant anti-EV-A71 activity in Vero cells.<sup>145</sup> Ursolic acid is a natural triterpene carboxylic acid compound with the therapeutic potential to prevent and combat EV-A71 infection.<sup>146,147</sup> Glycyrrhizic acid blocks the replication phase of EV-A71.<sup>148</sup> Corilagin dose-dependently reduces the cytopathic effect induced by EV-A71.149 Honeysuckle is the dried bud or firstblooming honeysuckle flower with antiviral activity. It attenuates EV-A71 infection by upregulating let-7a expression and targeting the viral genome.<sup>150</sup> However, no information on clinical trials of the above natural products against viral infections has been found.

### Monoclonal Antibody

After the virus invades the host, cells secrete specific antibodies to neutralize the virus particles, thereby treating viral infection. Monoclonal antibody (mAb) therapy is passive immunity and can effectively prevent infection by specific pathogens. Ideally, mAbs can neutralize not only the antiviral effect of a specific pathogen but also most subgenotypes of a specific pathogen species.<sup>151</sup> A number of mAbs for the treatment of viral infections are currently in clinical development. mAbs (3A12 and 2A10) exhibit significant inhibition of EV-A71 3D polymerase activity.<sup>152</sup> D5, H7, and C4 are mAbs targeting the VP1 GH loop. They do inhibit not only viral entry but also attachment, internalization, uncoating, and RNA release.<sup>153</sup> mAbs 51, 22, and 24 effectively neutralize EV-A71 particles and protect host cells.<sup>154,155</sup> Additionally, JL2 is an mAb that binds to hSCARB2. It plays an important role in protecting target cells from EV-A71 infection.<sup>156</sup>

#### **RNA** Interference

RNA interference (RNAi) is an effective means to target specific genes. RNAi therapies are used to treat infections caused by a variety of viruses. RNAi can interfere with EV-A71 genome replication. Small interfering RNAs (siRNAs) that are chemically synthesized can target different parts of the *EV-A71* gene for targeted inhibition. RNAi therapy is currently an effective method used to inhibit EV-A71 activity.<sup>13</sup> si-2C, si-3' untranslated region (UTR), and si-5' UTR have an anti–EV-A71 function. They can dosedependently block the transcription, translation, and replication of the EV-A71 virus.<sup>157,158</sup> siRNA-69, siRNA-294, and siRNA-319 are

3 siRNAs targeting the 2Apro region. They significantly reduce RNA replication.<sup>159</sup> The siRNA targeting 3Cpro and 3Dpol regions can improve survival in a mouse model of infection.<sup>89</sup> Furthermore, microR-9-5p not only modulates the levels of inflammatory factors induced by EV-A71 after attacking cells but also reduces the expression of VP1.<sup>160</sup>

Natural products are abundant. The medicinal effects of some natural products or herbal medicines have been proven after thousands of years of usage. Therefore, the research on antivirals could focus more on those with a convincing track record of use in Chinese medicine. Among them, the multiple targets targeted by natural ingredients are significant advantages for their antiviral effects. mAbs are indicated for patients who have developed resistance to habitual drug use and have used multiple drugs, but viral infection cannot be effectively controlled. RNAi is well targeted, but its delivery system may be potentially toxic in nonclinical toxicology studies in primates.<sup>71,136</sup> We may combine mAbs or RNAi with antivirals to enhance antiviral efficacy and achieve higher functional cure rates. The disease duration caused by EV-A71 infection is short, clinical symptoms are complex, and the effectiveness of drugs in clinical trials needs to be more readily determined, making developing antiviral agents verv challenging.

#### Vaccine

Owing to the lack of effective management and treatment for both mild and fatal EV-A71 infection, vaccination represents the best approach for controlling this life-threatening infection. In December 2015, the China Food and Drug Administration approved the first inactivated EV-A71 whole-virus vaccine for preventing severe HFMD. Different EV-A71 vaccines have been developed, such as inactivated virus vaccines and virus-like particle vaccines, DNA vaccines, subunit vaccines, and live attenuated vaccines.<sup>3</sup> From the previous study, activated whole-virus EV-A71 vaccines showed more effective when compared with other vaccines.<sup>161</sup> Most of the vaccine candidates are still at the preclinical stage of development, and currently a few inactivated EV71 vaccine candidates have completed clinical trials and 3 of them are approved by the China Food and Drug Administration.<sup>162</sup> All vaccine approaches and technologies have advantages and disadvantages, and the conventional EV-A71 vaccines such as live attenuated vaccines were limited by their safety concerns.<sup>163</sup> Modern approaches based on recombinant DNA technology, including recombinant subunit, virus-like particle, epitope-based, DNA, and live vector-based vaccines have been used to address the drawbacks and limitations of conventional vaccines.<sup>163,164</sup> However, improved effectiveness and broader protection against HFMD of these vaccines are needed to be further researched. Similarly to DNA vaccines, they demonstrated very weak and less potent immune responses in larger primates.<sup>11</sup> Hence, further studies on EV71 DNA vaccine candidates need to be conducted to improve their immunogenicity and protective efficacy. Furthermore, there are still a few challenges facing the worldwide use of EV71 vaccines, including the applicability against various EV-A71 pandemic strains in other countries, international requirements on vaccine production and quality control, standardization and harmonization on different pathogen-monitoring and -detecting methods, and so on. Therefore, with EV-A71 vaccines commercially available, there is still a long way to go before reaching effective protection against severe HFMD.

## Discussion

HFMD caused by EV-A71 threatens the health of infants and young children worldwide. Although fatal case studies of EV-A71 infection have been reported, the underlying mechanisms of EV-A71 infection, such as how the virus enters the CNS and causes pulmonary edema, have yet to be fully elucidated. To date, there is no approved specific and highly effective antiviral drug for HFMD caused by EV-A71. Therefore, we summarized the pathogenic mechanisms of EV-A71 infection and discussed the recent advances in inhibitors targeting EV-A71 entry, translation, polyprotein processing, replication, assembly, and release of new virions. Additionally, other inhibitors that inhibit EV-A71 through other pathways were also introduced, and the roles of each EV-A71 inhibitor in the virus life cycle were discussed. Although some anti-EV-A71 drugs have been discovered, developing anti--EV-A71 drugs still faces some difficulties, such as drug resistance, off-target effects, and drug safety. With the development of technology and multidisciplinary collaborations, we can gradually solve these problems that limit the research and development of agents by finding more suitable drug targets and constructing reasonable animal models to develop efficient and safe anti-EV-A71 drugs and provide specific and effective protection to children. Additionally, it is hoped that this review will provide guidance and ideas for researchers to develop anti-EV-A71 agents.

Patients with severe HFMD caused by EV-A71 infection are often treated symptomatically. For example, when patients develop intracranial hypertension complications, mannitol is used clinically to lower cranial pressure, and furosemide is added to relieve symptoms when necessary.<sup>165</sup> Blood biochemistry tests revealed elevated blood glucose levels in critically ill patients, and insulin may be applied in cases of severe hyperglycemia.<sup>166</sup> Clinical studies of inflammatory factors in the blood and cerebrospinal fluid of children with HFMD found that the white blood cell count and the level of inflammatory cytokines were significantly elevated in severe cases, and it was proposed that the inflammatory factor storm is one of the mechanisms of brain damage in severely ill children.<sup>3</sup> Therefore, glucocorticoid therapy, such as dexamethasone, is usually applied as appropriate, within 2 to 3 days. Administration of intravenous immunoglobulin and milrinone, a phosphodiesterase inhibitor, has been shown to modulate inflammation, reduce sympathetic overactivity, and improve survival in patients with EV-A71 autonomic nervous system dysregulation and pulmonary edema. Of course, physical adjuvant therapy will be commonly used, such as timely tracheal intubation and the use of positive pressure mechanical ventilation in case of respiratory dysfunction.<sup>167</sup>

EV-A71 has been recognized as highly neurotropic and associated with a diverse range of neurologic diseases, such as aseptic meningitis, brainstem encephalitis, encephalomyelitis, acute flaccid paralysis, and postinfectious neurologic syndromes. The pathogenesis of EV-A71-induced neurologic complications is caused by host-virus interaction, including direct damage by the virus and indirect injury mediated by immune and inflammatory responses.<sup>27</sup> Both innate and adaptive immune mechanisms are important for host defense against viral infections. The production of inflammatory cytokines and chemokines is a unique aspect of the immune responses in the CNS and systemic compartment to EV-A71 infection. Cytokines and chemokines released by EV-A71infected immune cells may contribute directly or indirectly to disease severity.<sup>25,27</sup> In addition, cellular signaling plays a critical role in the regulation of host innate immune and inflammatory pathogenesis.<sup>168</sup> Understanding the mechanism of cell signal transduction in regulating host cellular immunity and the occurrence of severe EV-A71 infection will not only help uncover the potential mechanisms of EV-A71 infection—induced pathogenesis but also provide clues for the design of therapeutic strategies against EV-A71 infection.

Given the high preventability of EV-A71 infection, we should actively take control measures to reduce its harm. The EV-A71 inactivated vaccine plays a vital role in the fight against severe HFMD outbreaks caused by EV-A71 infection. However, the coverage of monovalent vaccines is narrow and not universal. Moreover, the development of new combined multivalent vaccines has become complex owing to the diversity of serotypes, high cost, and lack of animal models. So, finding potent anti–EV-A71 drugs combined with vaccines is critical for eradicating EV-A71–associated diseases.

Recently, some drugs targeting EV-A71 have been discovered. Lactoferrin blocked EV-A71 entry by binding to the VP1 region.<sup>58</sup> Quercetin inhibited the activity of EV-A71 3Cpro.<sup>64,90</sup> They both target viruses. Currently, the EV-A71-targeting agents dominate the landscape. However, EV-A71 is an RNA virus, which has posed the problem of resistance owing to the rapid mutations and recombination events of EV-A71. Therefore, drugs with different targets and antiviral mechanisms are needed. The candidates with host-targeting mechanisms have been increasing in recent years. The bioactive compounds that target the host have broad biological and pharmacologic activities. They may have a more comprehensive range of antivirus activity spectrum. The common broadspectrum antiviral drugs targeting the host are IFNs. Although drugs targeting host cells may have a broader range of antivirus activity spectrum, they may be more toxic to the host and have offtarget effects. Therefore, there is an urgent need to develop specific and nontoxic antivirals. Natural products have abundant sources, low prices, stable efficacy, and low toxicity. They have been used extensively by researchers in the development of drugs. Natural products have been widely recognized in the treatment of common infectious diseases, such as influenza, infectious hepatitis, and enterovirus.144,169 However, the development of anti-EV-A71 agents has been restricted owing to the lack of advanced technology to extract active ingredients from natural products. It is believed that with the deepening of research, natural products will dominate in the field of antiviral treatment.

This review also discussed mAbs and RNAi. mAbs, a type of biological targeted therapy drug, have high specificity, good tolerance, and high safety. However, the purification process of mAbs is complicated, time consuming, labor intensive, and expensive. Moreover, the technologies of large-scale cell culture and animal cell antibody expression are yet to be overcome.<sup>170</sup> RNAi provides a promising antiviral treatment pathway. It can aid in the screening and development of new antivirals and enrich the information library of newly synthesized compounds on their mode of action. However, transferring siRNAs to the appropriate tissues at the appropriate time is a challenge in gene therapy.<sup>171</sup> As we gain more insights into the mechanisms, the knockdown technology might improve vastly with better-designed plasmidor virus-based vectors.

This review is a summary of the latest progress of existing anti–EV-A71 agents and discoveries, some with the potential for clinical development. Suramin has been used in clinical practice for decades, and its safety is not a concern. Suramin is currently in phase I of the HMFD clinical trial study and its antiviral activity is most worth tapping.<sup>48,49</sup> In China, no severe adverse drug reactions have occurred in Danshen pill injection in the past 8 years, MLB being the main component of Danshen pill injection. So, MLB has greater clinical potential.<sup>70,71</sup> There is a lot of research on inhibitors targeting viral proteins. However, there is no clinical information on most of the drugs. Among them, quercetin is currently in phase III of the COVID-19 clinical trial and has the greatest potential for anti-EV71 clinical development.<sup>89,90</sup> ITZ, a 3A inhibitor, is currently in phase I of clinical trials of RSV. RSV belongs to the same group of RNA viruses as EV71, so ITZ anti-EV71 has some potential for clinical development.<sup>80</sup> Additionally, Ye et al<sup>106</sup> showed that GS-5734 exhibited vigorous antiviral activity against EV-A71. GS-5734 has been approved for the treatment of severe acute respiratory syndrome-related coronavirus 2 infections and has high potential for clinical development among these drugs that inhibit EV-A71 genome replication. Minocycline is approved for marketing as an antibiotic and resveratrol is approved for anti-inflammatory/anticancer properties. They have a proven safety profile and have the potential to be developed into a clinical anti-EV71 drug.<sup>113,120</sup> Furthermore, berberine anti-COVID-19 studies are in phase IV of clinical trials with a proven safety profile. Researchers can prioritize berberine as an anti-EV71 candidate over other PI3K/Akt inhibitors.<sup>122</sup> In addition, many of the inhibitors of virion assembly and release and other EV-A71 inhibitors summarized in this study also have good anti-EV-A71 activity. However, there is a lack of information on clinical trials of these drugs against viral infections.

Researchers have been working hard to study the infection and host defense mechanisms of EV-A71 but have not fully elucidated the details of the EV-A71 transmission process and pathology. Appropriate animal models are needed to understand EV-A71 pathogenesis better and to develop highly effective antivirals. There have been reports of animal models of neonatal mice infected with mouse-adapted EV-A71 strains, cynomolgus monkeys, IFN receptor-deficient mice, transgenic mice with hSCARB2, and human PSGL-1 transgenic mice. However, it is difficult to establish an efficient infection model for EV-A71 owing to ethical, economic, and technical problems. The lack of animal models is also a reason that limits the development of drugs.<sup>172,173</sup> Additionally, drug development is a cumbersome process, a long cycle, and closely related to the application of technology. Approved drugs already have extensive clinical trial data and experience. Therefore, initial studies could focus on previously approved compounds, which can save more cost and time.

Finally, with the rapid development of science and technology and the continuous deepening of research, people will have a deeper understanding and insight into the virus life cycle. Additionally, researchers should closely search for relevant targets of antivirals and improve the methods of drug screening, isolation, and design, and actively carry out interdisciplinary cooperation. Thus, a new chapter is opened for the research and development of anti–EV-A71 drugs.

# Author Contributions

Y.W., K.H., and J.W. conceptualized the study. Y.W. and HH.L. wrote and prepared the original draft. Y.W., K.H., and J.W. reviewed, revised and edited the manuscript. K.H. and J.W. supervised the study. Y.W. and K.H. carried out project administration. Y.W., K.H., and J.W. carried out funding acquisition. D.H., Q.H., C.Y., and HL.L. provided comments and suggestions.

#### Data Availability

Not applicable.

# Funding

This research was funded by the Wuhan Knowledge Innovation Special Basic Research Project (2023020201010118) and the Open Project Funding of the Key Laboratory of Fermentation Engineering (Ministry of Education) (202209FE06) to Y.W.; the Key R&D Project of Hubei Province (Social Development), China (2022BCA018), and the Cooperative Innovation Center of Industrial Fermentation (Ministry of Education & Hubei Province), China (2022KF16), to K.H.; and the National Natural Science Foundation, China (81700013) to J.W.

#### Declaration of Competing Interest

The authors declare no conflict of interest.

#### Ethics Approval and Consent to Participate

Not applicable.

#### References

- Li XW, Ni X, Qian SY, et al. Chinese guidelines for the diagnosis and treatment of hand, foot and mouth disease (2018 edition). World J Pediatr. 2018;14(5):437–447.
- Nayak G, Bhuyan SK, Bhuyan R, Sahu A, Kar D, Kuanar A. Global emergence of enterovirus 71: a systematic review. *Beni Suef Univ J Basic Appl Sci.* 2022;11(1):78.
- Bello AM, Roshorm YM. Recent progress and advances towards developing enterovirus 71 vaccines for effective protection against human hand, foot and mouth disease (HFMD). *Biologicals*. 2022;79:1–9.
- Chong P, Liu CC, Chow YH, Chou AH, Klein M. Review of enterovirus 71 vaccines. *Clin Infect Dis.* 2015;60(5):797–803.
- Neumayr G, Pfister R, Mitterbauer G, et al. The emergence of enterovirus 71 as a major cause of acute neurological disease in young children of the Asia-Pacific region. J Pediatr Infect. 2006;1(1):17–23.
- Wang SM, Ho TS, Lin HC, Lei HY, Wang JR, Liu CC. Reemerging of enterovirus 71 in Taiwan: the age impact on disease severity. *Eur J Clin Microbiol Infect.* 2012;31(6):1219–1224.
- Wang SM, Liu CC. Enterovirus 71: epidemiology, pathogenesis and management. Expert Rev Anti Infect Ther. 2009;7(6):735–742.
- Ang LW, Phoon MC, Wu Y, Cutter J, James L, Chow VT. The changing seroepidemiology of enterovirus 71 infection among children and adolescents in Singapore. *BMC Infect Dis.* 2011;11:270.
- 9. Donato C, Le Hoi T, Hoa NT, et al. Genetic characterization of enterovirus 71 strains circulating in Vietnam in 2012. *Virology*. 2016;495:1–9.
- Takechi M, Fukushima W, Nakano T, et al. Nationwide survey of pediatric inpatients with hand, foot, and mouth disease, herpangina, and associated complications during an epidemic period in Japan: estimated number of hospitalized patients and factors associated with severe cases. J Epidemiol. 2019;29(9):354–362.
- 11. Chang LY. Enterovirus 71 in Taiwan. Pediatr Neonatol. 2008;49(4):103–112.
- Zeng M, El Khatib NF, Tu S, et al. Seroepidemiology of enterovirus 71 infection prior to the 2011 season in children in Shanghai. J Clin Virol. 2012;53(4):285–289.
- Shang L, Xu M, Yin Z. Antiviral drug discovery for the treatment of enterovirus 71 infections. *Antiviral Res.* 2013;97(2):183–194.
- Ang PY, Chong CWH, Alonso S. Viral determinants that drive enterovirus-A71 fitness and virulence. *Emerg Microbes Infect*. 2021;10(1):713–724.
- Wang H, Li Y. Recent progress on functional genomics research of enterovirus 71. Virol Sin. 2019;34(1):9–21.
- Baggen J, Thibaut HJ, Strating JRPM, van Kuppeveld FJM. The life cycle of non-polio enteroviruses and how to target it. *Nat Rev Microbiol*. 2018;16(6): 368–381.
- Ong KC, Badmanathan M, Devi S, Leong KL, Cardosa MJ, Wong KT. Pathologic characterization of a murine model of human enterovirus 71 encephalomyelitis. J Neuropathol Exp Neurol. 2008;67(6):532–542.
- Li R, Zou Q, Chen L, Zhang H, Wang Y. Molecular analysis of virulent determinants of enterovirus 71. *PLoS One*. 2011;6(10):e26237.
- 19. Huang HI, Weng KF, Shih SR. Viral and host factors that contribute to pathogenicity of enterovirus 71. *Future Microbiol*. 2012;7(4):467–479.

- Solomon T, Lewthwaite P, Perera D, Cardosa MJ, McMinn P, Ooi MH. Virology, epidemiology, pathogenesis, and control of enterovirus 71. *Lancet Infect Dis.* 2010;10(11):778–790.
- 21. Gao L, Lin P, Liu S, et al. Pathological examinations of an enterovirus 71 infection: an autopsy case. Int J Clin Exp Pathol. 2014;7(8):5236–5241.
- 22. Chang LY, Chang IS, Chen WJ, et al. HLA-A33 is associated with susceptibility to enterovirus 71 infection. *Pediatrics*. 2008;122(6):1271–1276.
- Wang SM, Chen IC, Liao YT, Liu CC. The clinical correlation of regulatory T cells and cyclic adenosine monophosphate in enterovirus 71 infection. *PLoS One.* 2014;9(7):e102025.
- 24. Pathinayake PS, Hsu AC, Wark PA. Innate immunity and immune evasion by enterovirus 71. *Viruses*. 2015;7(12):6613–6630.
- Zhang W, Huang Z, Huang M, Zeng J. Predicting severe enterovirus 71infected hand, foot, and mouth disease: cytokines and chemokines. *Mediators Inflamm*. 2020;2020:9273241.
- Lee KY. Enterovirus 71 infection and neurological complications. Korean J Pediatr. 2016;59(10):395–401.
- Wang SM, Lei HY, Liu CC. Cytokine immunopathogenesis of enterovirus 71 brain stem encephalitis. Clin Dev Immunol. 2012;2012:876241.
- Tu YF, Lin CH, Lee HT, et al. Elevated cerebrospinal fluid endothelin 1 associated with neurogenic pulmonary edema in children with enterovirus 71 encephalitis. *Int J Infect Dis.* 2015;34:105–111.
   Tantsis EM, Prelog K, Alper G, et al. Magnetic resonance imaging in
- 29. Tantsis EM, Prelog K, Alper G, et al. Magnetic resonance imaging in enterovirus-71, myelin oligodendrocyte glycoprotein antibody, aquaporin-4 antibody, and multiple sclerosis-associated myelitis in children. *Dev Med Child Neurol.* 2019;61(9):1108–1116.
- **30.** Long L, Xu L, Xiao Z, et al. Neurological complications and risk factors of cardiopulmonary failure of EV-A71-related hand, foot and mouth disease. *Sci Rep.* 2016;6:23444.
- Shen WC, Chiu HH, Chow KC, Tsai CH. MR imaging findings of enteroviral encephaloymelitis: an outbreak in Taiwan. *AJNR Am J Neuroradiol*. 1999;20(10):1889–1895.
- **32.** Yu P, Gao Z, Zong Y, et al. Distribution of enterovirus 71 RNA in inflammatory cells infiltrating different tissues in fatal cases of hand, foot, and mouth disease. *Arch Virol.* 2015;160(1):81–90.
- Xing J, Wang K, Wang G, Li N, Zhang Y. Recent advances in enterovirus A71 pathogenesis: a focus on fatal human enterovirus A71 infection. *Arch Virol.* 2022;167(12):2483–2501.
- Dang M, Wang X, Wang Q, et al. Molecular mechanism of SCARB2-mediated attachment and uncoating of EV71. *Protein Cell*. 2014;5(9):692–703.
- Yamayoshi S, Yamashita Y, Li J, et al. Scavenger receptor B2 is a cellular receptor for enterovirus 71. Nat Med. 2009;15(7):798–801.
- Nishimura Y, Shimojima M, Tano Y, Miyamura T, Wakita T, Shimizu H. Human P-selectin glycoprotein ligand-1 is a functional receptor for enterovirus 71. Nat Med. 2009;15(7):794–797.
- **37.** Tan CW, Poh CL, Sam IC, Chan YF. Enterovirus 71 uses cell surface heparan sulfate glycosaminoglycan as an attachment receptor. *J Virol.* 2013;87(1): 611–620.
- Yang B, Chuang H, Yang KD. Sialylated glycans as receptor and inhibitor of enterovirus 71 infection to DLD-1 intestinal cells. *Virol J.* 2009;6:141.
- Du N, Cong H, Tian H, et al. Cell surface vimentin is an attachment receptor for enterovirus 71. J Virol. 2014;88(10):5816–5833.
- Su PY, Wang YF, Huang SW, et al. Cell surface nucleolin facilitates enterovirus 71 binding and infection. J Virol. 2015;89(8):4527-4538.
- Yang SL, Chou YT, Wu CN, Ho MS. Annexin II binds to capsid protein VP1 of enterovirus 71 and enhances viral infectivity. J Virol. 2011;85(22): 11809–11820.
- 42. Qing J, Wang Y, Sun Y, et al. Cyclophilin A associates with enterovirus-71 virus capsid and plays an essential role in viral infection as an uncoating regulator. *PLoS Pathog.* 2014;10(10):e1004422.
- **43.** Yeung ML, Jia L, Yip CCY, et al. Human tryptophanyl-tRNA synthetase is an IFN-gamma-inducible entry factor for enterovirus. *J Clin Invest.* 2018;128(11):5163–5177.
- 44. Wang RY, Kuo RL, Ma WC, et al. Heat shock protein-90-beta facilitates enterovirus 71 viral particles assembly. *Virology*. 2013;443(2):236–247.
- **45.** He QQ, Ren S, Xia ZC, Cheng ZK, Peng NF, Zhu Y. Fibronectin facilitates enterovirus 71 infection by mediating viral entry. *J Virol.* 2018;92(9). e02251-17.
- 46. Too IHK, Bonne I, Tan EL, Chu JJH, Alonso S. Prohibitin plays a critical role in enterovirus 71 neuropathogenesis. *PLoS Pathog.* 2018;14(1), e1006778.
- Wang Y, Qing J, Sun Y, Rao Z. Suramin inhibits EV71 infection. Antiviral Res. 2014;103:1–6.
- **48.** Ord JJ, Streeter E, Jones A, et al. Phase I trial of intravesical Suramin in recurrent superficial transitional cell bladder carcinoma. *Br J Cancer*. 2005;92(12):2140–2147.
- 49. Laterra JJ, Grossman SA, Carson KA, et al. Suramin and radiotherapy in newly diagnosed glioblastoma: phase 2 NABTT CNS Consortium study. *Neuro Oncol.* 2004;6(1):15–20.
- Arita M, Wakita T, Shimizu H. Characterization of pharmacologically active compounds that inhibit poliovirus and enterovirus 71 infectivity. J Gen Virol. 2008;89(10):2518–2530.
- **51.** Yue Y, Li Z, Li P, et al. Antiviral activity of a polysaccharide from Laminaria japonica against enterovirus 71. *Biomed Pharmacother*. 2017;96:256–262.

- Li Z, Cui B, Liu X, et al. Virucidal activity and the antiviral mechanism of acidic polysaccharides against enterovirus 71 infection in vitro. *Microbiol Immunol*. 2020;64(3):189–201.
- Uddin MB, Lee BH, Nikapitiya C, et al. Inhibitory effects of bee venom and its components against viruses in vitro and in vivo. J Microbiol. 2016;54(12): 853–866.
- Masomian M, Lalani S, Poh CL. Molecular docking of SP40 peptide towards cellular receptors for enterovirus 71 (EV-A71). *Molecules*. 2021;26(21):6576.
- Ren G, Ding G, Zhang H, et al. Antiviral activity of sophoridine against enterovirus 71 in vitro. J Ethnopharmacol. 2019;236:124–128.
- Meng T, Jia Q, Wong SM, Chua KB. In vitro and in vivo inhibition of the infectivity of human enterovirus 71 by a sulfonated food azo dye, brilliant black BN. J Virol. 2019;93(17):e00061-19.
- 57. Yuan M, Yan J, Xun J, et al. Enhanced human enterovirus 71 infection by endocytosis inhibitors reveals multiple entry pathways by enterovirus causing hand-foot-and-mouth diseases. *Virol J.* 2018;15(1):1.
- Weng TY, Chen LC, Shyu HW, et al. Lactoferrin inhibits enterovirus 71 infection by binding to VP1 protein and host cells. *Antiviral Res.* 2005;67(1):31–37.
- Wang X, Peng W, Ren J, et al. A sensor-adaptor mechanism for enterovirus uncoating from structures of EV71. Nat Struct Mol Biol. 2012;19(4):424–429.
- 60. Li P, Yu J, Hao F, et al. Discovery of potent EV71 capsid inhibitors for treatment of HFMD. ACS Med Chem Lett. 2017;8(8):841-846.
- 61. Zhang M, Wang Y, He W, et al. Design, synthesis, and evaluation of novel enterovirus 71 inhibitors as therapeutic drug leads for the treatment of human hand, foot, and mouth disease. *I Med Chem.* 2020;63(3):1233–1244.
- 62. Lin WY, Yu YJ, Jinn TR. Evaluation of the virucidal effects of rosmarinic acid against enterovirus 71 infection via in vitro and in vivo study. *Virol J.* 2019;16(1):94.
- **63.** Yan W, Qing J, Mei H, et al. Identification, synthesis and pharmacological evaluation of novel anti-EV71 agents via cyclophilin A inhibition. *Bioorg Med Chem Lett.* 2015;25(24):5682–5686.
- 64. Tijsma A, Franco D, Tucker S, et al. The capsid binder Vapendavir and the novel protease inhibitor SG85 inhibit enterovirus 71 replication. *Antimicrob Agents Chemother*. 2014;58(11):6990–6992.
- 65. Wang M, Jiang S, Wang Y. Recombinant VP1 protein expressed in Pichia pastoris induces protective immune responses against EV71 in mice. *Biochem Biophys Res Commun*. 2013;430(1):387–393.
- 66. Davila-Calderon J, Patwardhan NN, Chiu LY, et al. IRES-targeting small molecule inhibits enterovirus 71 replication via allosteric stabilization of a ternary complex. *Nat Commun.* 2020;11(1):4775.
- 67. Chung YC, Hsieh FC, Lin YJ, et al. Magnesium lithospermate B and rosmarinic acid, two compounds present in Salvia miltiorrhiza, have potent antiviral activity against enterovirus 71 infections. *Eur J Pharmacol.* 2015;755:127–133.
- **68.** Wang J, Du J, Wu Z, Jin Q. Quinacrine impairs enterovirus 71 RNA replication by preventing binding of polypyrimidine-tract binding protein with internal ribosome entry sites. *PLoS One.* 2013;8(1):e52954.
- 69. Tsai FJ, Lin CW, Lai CC, et al. Kaempferol inhibits enterovirus 71 replication and internal ribosome entry site (IRES) activity through FUBP and HNRP proteins. *Food Chem.* 2011;128(2):312–322.
- Yan YY, Yang YH, Wang WW, et al. Post-marketing safety surveillance of the Salvia Miltiorrhiza depside salt for infusion: a real world study. *PLoS One*. 2017;12(1):e0170182.
- Chaudhuri S, Symons JA, Deval J. Innovation and trends in the development and approval of antiviral medicines: 1987-2017 and beyond. *Antiviral Res.* 2018;155:76–88.
- Mu Z, Wang B, Zhang X, et al. Crystal structure of 2A proteinase from hand, foot and mouth disease virus. J Mol Biol. 2013;425(22):4530–4543.
- 73. Lalani S, Gew LT, Poh CL. Antiviral peptides against enterovirus A71 causing hand, foot and mouth disease. *Peptides*. 2021;136:170443.
- Falah N, Montserret R, Lelogeais V, et al. Blocking human enterovirus 71 replication by targeting viral 2A protease. J Antimicrob Chemother. 2012;67(12):2865–2869.
- Wang CY, Huang AC, Hour MJ, et al. Antiviral potential of a novel compound CW-33 against enterovirus A71 via inhibition of viral 2A protease. *Viruses*. 2015;7(6):3155–3171.
- Xue F, Luo X, Ye C, Ye W, Wang Y. Inhibitory properties of 2-substituent-1Hbenzimidazole-4-carboxamide derivatives against enteroviruses. *Bioorg Med Chem.* 2011;19(8):2641–2649.
- Tang WF, Yang SY, Wu BW, et al. Reticulon 3 binds the 2C protein of enterovirus 71 and is required for viral replication. *J Biol Chem.* 2007;282(8): 5888–5898.
- Xiao X, Lei X, Zhang Z, et al. Enterovirus 3A facilitates viral replication by promoting phosphatidylinositol 4-kinase IIIbeta-ACBD3 interaction. J Virol. 2017;91(19):e00791-17.
- **79.** Arita M, Takebe Y, Wakita T, Shimizu H. A bifunctional anti-enterovirus compound that inhibits replication and the early stage of enterovirus 71 infection. *J Gen Virol.* 2010;91(11):2734–2744.
- Gao Q, Yuan S, Zhang C, et al. Discovery of itraconazole with broadspectrum in vitro antienterovirus activity that targets nonstructural protein 3A. Antimicrob Agents Chemother. 2015;59(5):2654–2665.
- **81.** Tang F, Xia H, Wang P, et al. The identification and characterization of nucleic acid chaperone activity of human enterovirus 71 nonstructural protein 3AB. *Virology*. 2014;464-465:353–364.

- Velu AB, Chen GW, Hsieh PT, et al. BPR-3P0128 inhibits RNA-dependent RNA polymerase elongation and VPg uridylylation activities of enterovirus 71. Antiviral Res. 2014;112:18–25.
- Weng KF, Li ML, Hung CT, Shih SR. Enterovirus 71 3C protease cleaves a novel target CstF-64 and inhibits cellular polyadenylation. *PLoS Pathog.* 2009;5(9):e1000593.
- **84.** Zhang X, Song Z, Qin B, et al. Rupintrivir is a promising candidate for treating severe cases of enterovirus-71 infection: evaluation of antiviral efficacy in a murine infection model. *Antiviral Res.* 2013;97(3):264–269.
- **85.** Dai W, Jochmans D, Xie H, et al. Design, synthesis, and biological evaluation of peptidomimetic aldehydes as broad-spectrum inhibitors against enterovirus and SARS-CoV-2. *J Med Chem.* 2022;65(4): 2794–2808.
- Wang Y, Cao L, Zhai Y, et al. Inhibition of enterovirus 71 replication by an alpha-hydroxy-nitrile derivative NK-1.9k. Antiviral Res. 2017;141:91–100.
- 87. Wang Y, Yang B, Zhai Y, Yin Z, Sun Y, Rao Z. Peptidyl aldehyde NK-1.8k suppresses enterovirus 71 and enterovirus 68 infection by targeting protease 3C. Antimicrob Agents Chemother. 2015;59(5):2636–2646.
- 88. Tan J, George S, Kusov Y, et al. 3C protease of enterovirus 68: structurebased design of Michael acceptor inhibitors and their broad-spectrum antiviral effects against picornaviruses. J Virol. 2013;87(8):4339–4351.
- 89. Diarimalala RO, Hu M, Wei Y, Hu K. Recent advances of enterovirus 71 [Formula: see text] targeting inhibitors. *Virol J.* 2020;17(1):173.
- Yao C, Xi C, Hu K, et al. Inhibition of enterovirus 71 replication and viral 3C protease by quercetin. *Virol J.* 2018;15(1):116.
- **91.** Jiang H, Weng L, Zhang N, et al. Biochemical characterization of enterovirus 71 3D RNA polymerase. *Biochim Biophys Acta*. 2011;1809(3):211–219.
- **92.** Shang L, Wang Y, Qing J, et al. An adenosine nucleoside analogue NITD008 inhibits EV71 proliferation. *Antiviral Res.* 2014;112:47–58.
- Hung HC, Chen TC, Fang MY, et al. Inhibition of enterovirus 71 replication and the viral 3D polymerase by aurintricarboxylic acid. J Antimicrob Chemother. 2010;65(4):676–683.
- 94. van der Linden L, Vives-Adrian L, Selisko B, et al. The RNA template channel of the RNA-dependent RNA polymerase as a target for development of antiviral therapy of multiple genera within a virus family. *PLoS Pathog.* 2015;11(3):e1004733.
- Chen TC, Chang HY, Lin PF, et al. Novel antiviral agent DTriP-22 targets RNAdependent RNA polymerase of enterovirus 71. *Antimicrob Agents Chemother*. 2009;53(7):2740–2747.
- **96.** Stopa JD, Neuberg D, Puligandla M, Furie B, Flaumenhaft R, Zwicker JI. Protein disulfide isomerase inhibition blocks thrombin generation in humans by interfering with platelet factor V activation. *JCI Insight*. 2017;2(1):e89373.
- Solomon S. A practical guide to clinical virology. Yale J Biol Med. 1991;64(1): 104–105.
- Oxford JS, Whitley RJ. Antiviral drugs. In: Haaheim LR, Pattison JR, Whitley RJ, eds. A Practical Guide to Clinical Virology. 2nd Edition. John Wiley & Sons, Ltd; 2002:21–35.
- 99. van der Schaar HM, Leyssen P, Thibaut HJ, et al. A novel, broad-spectrum inhibitor of enterovirus replication that targets host cell factor phosphatidylinositol 4-kinase IIIβ. Antimicrob Agents Chemother. 2013;57(10): 4971–4981.
- 100. Zhang H, Tao L, Fu WW, et al. Prenylated benzoylphloroglucinols and xanthones from the leaves of *Garcinia oblongifolia* with antienteroviral activity. *J Nat Prod.* 2014;77(4):1037–1046.
- 101. Zhang G, Zhou F, Gu B, et al. In vitro and in vivo evaluation of ribavirin and pleconaril antiviral activity against enterovirus 71 infection. *Arch Virol.* 2012;157(4):669–679.
- **102.** Li ZH, Li CM, Ling P, et al. Ribavirin reduces mortality in enterovirus 71infected mice by decreasing viral replication. *J Infect Dis.* 2008;197(6): 854–857.
- **103.** Wei Y, Shi L, Wang K, et al. Discovery of gramine derivatives that inhibit the early stage of EV71 replication in vitro. *Molecules.* 2014;19(7): 8949–8964.
- 104. Yang Y, Xiu J, Liu J, et al. Chebulagic acid, a hydrolyzable tannin, exhibited antiviral activity in vitro and in vivo against human enterovirus 71. Int J Mol Sci. 2013;14(5):9618–9627.
- 105. Zhang W, Qiao H, Lv Y, et al. Apigenin inhibits enterovirus-71 infection by disrupting viral RNA association with trans-acting factors. *PLoS One*. 2014;9(10):e110429.
- 106. Ye W, Yao M, Dong Y, et al. Remdesivir (GS-5734) impedes enterovirus replication through viral RNA synthesis inhibition. *Front Microbiol*. 2020;11: 1105.
- **107.** Liu ML, Lee YP, Wang YF, et al. Type I interferons protect mice against enterovirus 71 infection. *J Gen Virol*. 2005;86(Pt 12):3263–3269.
- 108. Yi L, Lu J, Kung HF, He ML. The virology and developments toward control of human enterovirus 71. Crit Rev Microbiol. 2011;37(4):313–327.
- 109. Kang NX, Zou Y, Liang QH, et al. Anemoside B4 inhibits enterovirus 71 propagation in mice through upregulating 14-3-3 expression and type I interferon responses. Acta Pharmacol Sin. 2022;43(4):977–991.
- 110. Lin CW, Wu CF, Hsiao NW, et al. Aloe-emodin is an interferon-inducing agent with antiviral activity against Japanese encephalitis virus and enterovirus 71. Int J Antimicrob Agents. 2008;32(4):355–359.

- 111. Kim JH, Weeratunga P, Kim MS, et al. Inhibitory effects of an aqueous extract from Cortex Phellodendri on the growth and replication of broadspectrum of viruses in vitro and in vivo. BMC Complement Altern Med. 2016:16:265.
- 112. Li M, Yang X, Guan C, et al. Andrographolide sulfonate reduces mortality in enterovirus 71 infected mice by modulating immunity. *Int Immuno*pharmacol. 2018;55:142–150.
- 113. Liao YT, Wang SM, Chen SH. Anti-inflammatory and antiviral effects of minocycline in enterovirus 71 infections. Biomed Pharmacother. 2019;118: 109271
- 114. Wang YC, Yi TY, Lin KH. In vitro activity of Paris polyphylla smith against enterovirus 71 and coxsackievirus B3 and its immune modulation. Am J Chin Med. 2011;39(6):1219-1234.
- 115. Zhu L, Li W, Qi G, et al. The immune mechanism of intestinal tract toll-like receptor in mediating EV71 virus type severe hand-foot-and-mouth disease and the MAPK pathway. Exp Ther Med. 2017;13(5):2263-2266.
- 116. Zhang Z, Wang B, Wu S, et al. PD169316, a specific p38 inhibitor, shows antiviral activity against enterovirus71. Virology. 2017;508:150-158.
- 117. Wang H, Zhang D, Ge M, Li Z, Jiang J, Li Y. Formononetin inhibits enterovirus 71 replication by regulating COX-2/PGE(2) expression. Virol J. 2015;12:35.
- 118. Wang HQ, Hu J, Yan HY, Wu S, Li YH. Corydaline inhibits enterovirus 71 replication by regulating COX-2 expression. J Asian Nat Prod Res. 2017;19(11):1124-1133.
- 119. Wang C, Zhang H, Xu F, et al. Substituted 3-benzylcoumarins as allosteric MEK1 inhibitors: design, synthesis and biological evaluation as antiviral agents. Molecules. 2013;18(5):6057-6091.
- 120. Zhang L, Li Y, Gu Z, et al. Resveratrol inhibits enterovirus 71 replication and pro-inflammatory cytokine secretion in rhabdosarcoma cells through blocking IKKs/NF-κB signaling pathway. PLoS One. 2015;10(2): e0116879.
- 121. Zhang H, Li F, Pan Z, Wu Z, Wang Y, Cui Y. Activation of PI3K/Akt pathway limits JNK-mediated apoptosis during EV71 infection. Virus Res. 2014;192: 74-84.
- 122. Wang H, Li K, Ma L, et al. Berberine inhibits enterovirus 71 replication by downregulating the MEK/ERK signaling pathway and autophagy. Virol J. 2017;14(1):2.
- 123. Wang YX, Yang L, Wang HQ, et al. Synthesis and evolution of berberine derivatives as a new class of antiviral agents against enterovirus 71 through the MEK/ERK pathway and autophagy. Molecules. 2018;23(8):2084.
- 124. Li X, Liu Y, Wu T, et al. The antiviral effect of baicalin on enterovirus 71 in vitro. Viruses. 2015;7(8):4756-4771.
- 125. Zhang Q, Zhao B, Chen X, et al. GS-9620 inhibits enterovirus 71 replication mainly through the NF-kB and PI3K-AKT signaling pathways. Antiviral Res. 2018:153:39-48.
- 126. Guillin OM, Vindry C, Ohlmann T, Chavatte L. Selenium, selenoproteins and viral infection. *Nutrients*. 2019;11(9):2101.
- 127. Zhong J, Xia Y, Hua L, et al. Functionalized selenium nanoparticles enhance the anti-EV71 activity of oseltamivir in human astrocytoma cell model. Artif Cells Nanomed Biotechnol. 2019;47(1):3485–3491.
- 128. Ho HY, Cheng ML, Weng SF, Leu YL, Chiu DT. Antiviral effect of epigallocatechin gallate on enterovirus 71. J Agric Food Chem. 2009;57(14): 6140-6147.
- 129. Choi HJ, Song JH, Park KS, Baek SH. In vitro anti-enterovirus 71 activity of gallic acid from Woodfordia fruticosa flowers. Lett Appl Microbiol. 2010:50(4):438-440.
- 130. Lv X, Qiu M, Chen D, Zheng N, Jin Y, Wu Z. Apigenin inhibits enterovirus 71 replication through suppressing viral IRES activity and modulating cellular INK pathway. Antiviral Res. 2014;109:30–41.
- 131. Xu L, Su W, Jin J, et al. Identification of luteolin as enterovirus 71 and coxsackievirus A16 inhibitors through reporter viruses and cell viabilitybased screening. Viruses. 2014;6(7):2778-2795.
- 132. Cetin G, Klafack S, Studencka-Turski M, Kruger E, Ebstein F. The ubiquitinproteasome system in immune cells. Biomolecules. 2021;11(1):60.
- 133. Lin L. Oin Y. Wu H. et al. Pyrrolidine dithiocarbamate inhibits enteroyirus 71 replication by down-regulating ubiquitin-proteasome system. Virus Res. 2015:195:207-216.
- 134. Tsou YL, Lin YW, Chang HW, et al. Heat shock protein 90: role in enterovirus 71 entry and assembly and potential target for therapy. PLoS One. 2013;8(10):e77133.
- 135. Garrido-Mesa N, Zarzuelo A, Galvez J. Minocycline: far beyond an antibiotic. Br J Pharmacol. 2013;169(2):337-352.
- 136. Desai MC, Meanwell NA, Thurston DE. Successful Strategies for the Discovery of Antiviral Drugs. The Royal Society of Chemistry; 2013.
- 137. Tan YW, Hong WJ, Chu JJ. Inhibition of enterovirus VP4 myristoylation is a potential antiviral strategy for hand, foot and mouth disease. Antiviral Res. 2016:133:191-195.
- 138. Huang L, Yue J. The interplay of autophagy and enterovirus. Semin Cell Dev Biol. 2020;101:12-19.
- 139. Dai W, Wu Y, Bi J, et al. Antiviral effects of Retro-2cycl and Retro-2.1 against enterovirus 71 in vitro and in vivo. Antiviral Res. 2017;144:311-321.
- 140. Wang H, Guo T, Yang Y, Yu L, Pan X, Li Y. Lycorine derivative LY-55 inhibits EV71 and CVA16 replication through downregulating autophagy. Front Cell Infect Microbiol. 2019;9:277.

- 141. Won M, Jun EJ, Khim M, et al. Antiviral protection against enterovirus 71 mediated by autophagy induction following FLICE-inhibitory protein inactivation. Virus Res. 2012;169(1):316-320.
- 142. Talaei S, Mellatyar H, Asadi A, Akbarzadeh A, Sheervalilou R, Zarghami N. Spotlight on 17-AAG as an Hsp90 inhibitor for molecular targeted cancer treatment. *Chem Biol Drug Des.* 2019;93(5):760–786. 143. Wang L, Wang J, Wang L, Ma S, Liu Y. Anti-enterovirus 71 agents of natural
- products. Molecules. 2015;20(9):16320-16333.
- 144. Musarra-Pizzo M, Pennisi R, Ben-Amor I, Mandalari G, Sciortino MT. Antiviral activity exerted by natural products against human viruses. Viruses. 2021;13(5):828.
- 145. Choi HJ, Lim CH, Song JH, Baek SH, Kwon DH. Antiviral activity of raoulic acid from Raoulia australis against Picornaviruses. Phytomedicine. 2009;16(1):35-39.
- 146. Zhao CH, Xu J, Zhang YQ, Zhao LX, Feng B. Inhibition of human enterovirus 71 replication by pentacyclic triterpenes and their novel synthetic derivatives. Chem Pharm Bull (Tokyo). 2014;62(8):764-771.
- 147. Chiang LC, Ng LT, Cheng PW, Chiang W, Lin CC. Antiviral activities of extracts and selected pure constituents of Ocimum basilicum. Clin Exp Pharmacol Physiol. 2005;32(10):811-816.
- 148. Wang J, Chen X, Wang W, et al. Glycyrrhizic acid as the antiviral component of Glycyrrhiza uralensis Fisch. against coxsackievirus A16 and enterovirus 71 of hand foot and mouth disease. J Ethnopharmacol. 2013;147(1): 114-121.
- 149. Yeo SG, Song JH, Hong EH, et al. Antiviral effects of Phyllanthus urinaria containing corilagin against human enterovirus 71 and Coxsackievirus A16 in vitro. Arch Pharm Res. 2015;38(2):193-202.
- 150. Lee YR, Chang CM, Yeh YC, et al. Honeysuckle aqueous extracts induced let-7a suppress EV71 replication and pathogenesis in vitro and in vivo and is predicted to inhibit SARS-CoV-2. Viruses. 2021;13(2):208.
- 151. Rattanapisit K, Chao Z, Siriwattananon K, Huang Z, Phoolcharoen W. Plantproduced anti-enterovirus 71 (EV71) monoclonal antibody efficiently protects mice against EV71 infection. Plants (Basel). 2019;8(12):560.
- 152. Li Y, Yu J, Qi X, Yan H. Monoclonal antibody against EV71 3Dpol inhibits the polymerase activity of RdRp and virus replication. BMC Immunol. 2019;20(1):6.
- 153. Ku Z, Ye X, Shi J, Wang X, Liu Q, Huang Z. Single neutralizing monoclonal antibodies targeting the VP1 GH loop of enterovirus 71 inhibit both virus attachment and internalization during viral entry. J Virol. 2015;89(23): 12084-12095.
- 154. Lim XF, Jia Q, Khong WX, et al. Characterization of an isotype-dependent monoclonal antibody against linear neutralizing epitope effective for prophylaxis of enterovirus 71 infection. PLoS One. 2012;7(1):e29751.
- 155. Xu L, Huang KJ, Ho TS, et al. Monoclonal antibodies for diagnosis of enterovirus 71. Monoclon Antib Immunodiagn Immunother. 2013:32(6): 386-394.
- 156. Zhang X, Yang P, Wang N, et al. The binding of a monoclonal antibody to the apical region of SCARB2 blocks EV71 infection. Protein Cell. 2017;8(8): 590 - 600
- 157. Sim AC, Luhur A, Tan TM, Chow VT, Poh CL. RNA interference against enterovirus 71 infection. Virology. 2005;341(1):72-79.
- 158. Deng JX, Nie XJ, Lei YF, et al. The highly conserved 5' untranslated region as an effective target towards the inhibition of Enterovirus 71 replication by unmodified and appropriate 2'-modified siRNAs. J Biomed Sci. 2012;19(1): 73.
- 159. Liu H, Qin Y, Kong Z, et al. siRNA targeting the 2Apro genomic region prevents enterovirus 71 replication in vitro. PLos One. 2016;11(2):e0149470.
- **160.** Li B, Zheng J. MicroR-9-5p suppresses EV71 replication through targeting NFκB of the RIG-1-mediated innate immune response. *FEBS Open Bio.* 2018;8(9):1457-1470.
- 161. Zhu FC, Meng FY, Li JX, et al. Efficacy, safety, and immunology of an inactivated alum-adjuvant enterovirus 71 vaccines in children in China: a multicentre, randomized, double-blind, placebo-controlled, phase 3 trial. Lancet. 2013;381(9882):2024-2032.
- 162. Lu S. EV71 vaccines: a milestone in the history of global vaccine development. Emerg Microbes Infect. 2014;3(4):e27.
- 163. Lauring AS, Jones JO, Andino R. Rationalizing the development of live attenuated virus vaccines. Nat Biotechnol. 2010;28(6):573-579.
- 164. Sakurai A, Ogawa T, Matsumoto J, et al. Regulatory aspects of quality and safety for live recombinant viral vaccines against infectious diseases in Japan. Vaccine. 2019;37(43):6573-6579.
- 165. Wang SM. Milrinone in enterovirus 71 brain stem encephalitis. Front Pharmacol. 2016;7:82.
- 166. Li J, Lin A, Yu C, et al. Association of Enterovirus 71 encephalitis with the interleukin-8 gene region in Chinese children. Infect Dis (Lond). 2015;47(6): 418-422.
- 167. Hua Y, Wang Y, Gong W. Inflammatory cytokine profiles of serum and cerebrospinal fluid in Chinese children with hand, foot and mouth disease. Int I Clin Exp Pathol. 2017;10(11):11022-11029.
- 168. Jin Y, Zhang R, Wu W, Duan G. Antiviral and inflammatory cellular signaling associated with enterovirus 71 infection. Viruses. 2018;10(4):155.
- 169. Cary DC, Peterlin BM. Natural products and HIV/AIDS. AIDS Res Hum Retroviruses. 2018;34(1):31-38.

- Mould DR, Meibohm B. Drug development of therapeutic monoclonal an-tibodies. *BioDrugs*. 2016;30(4):275–293.
   Agrawal N, Dasaradhi PV, Mohmmed A, Malhotra P, Bhatnagar RK, Mukherjee SK. RNA interference: biology, mechanism, and applications. *Microbiol Mol Biol Rev*. 2003;67(4):657–685.
- 172. Yi EJ, Shin YJ, Kim JH, Kim TG, Chang SY. Enterovirus 71 infection and vaccines. *Clin Exp Vaccine Res.* 2017;6(1):4–14.
  173. Shih C, Liao CC, Chang YS, Wu SY, Chang CS, Liou AT. Immunocompetent and immunodeficient mouse models for enterovirus 71 pathogenesis and therapy. *Viruses.* 2018;10(12):674.